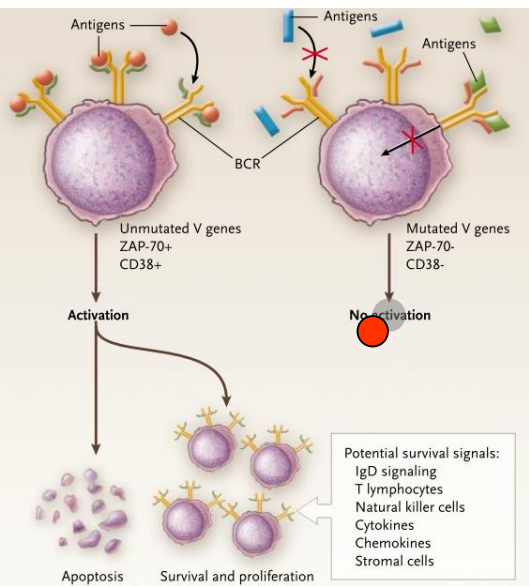
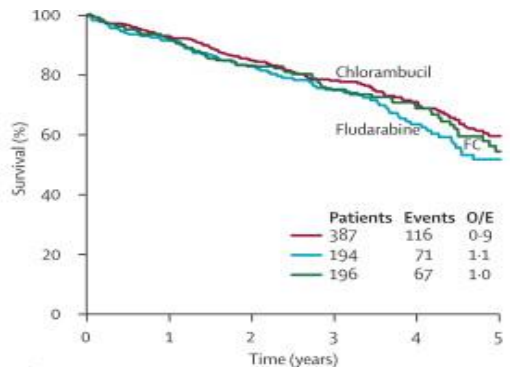


## Where did we start from

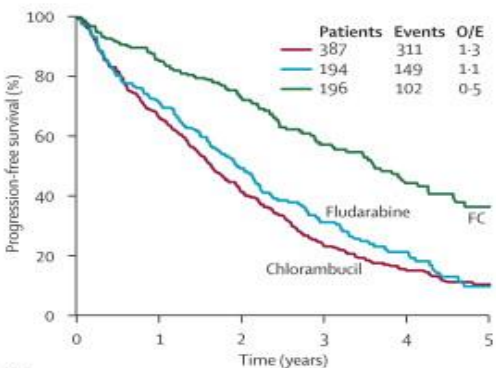
### Activation following BCR stimulation



### FC improves PFS, but not OS

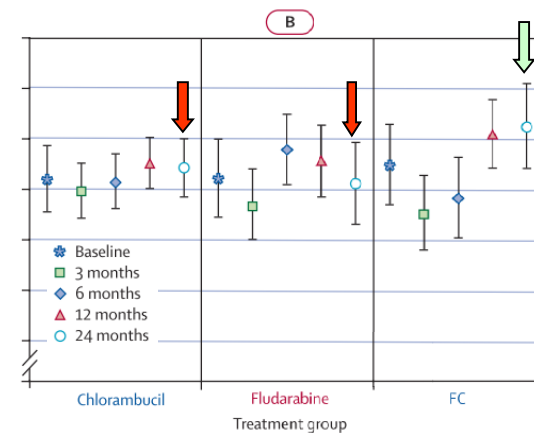


Patients at risk	0	1	2	3	4	5
Chlorambucil	387	359	302	201	132	60
Fludarabine	194	177	150	100	62	29
FC	196	181	149	97	70	30



Patients at risk	0	1	2	3	4	5
Chlorambucil	387	258	151	61	30	11
Fludarabine	194	139	91	40	21	5
FC	196	168	131	74	43	19

### Longer PFS = better QOL



Mechanisms of disease: Chronic Lymphocytic Leukemia  
Nicholas Chiorazzi, M.D., Kanti R. Rai, M.B., B.S., and Manlio Ferrarini, M.D.  
N Engl J Med 2005;352:804-15.

Chlor vs F vs FC in CLL LRF CLL4 trial  
Catovsky et al, Lancet 2007

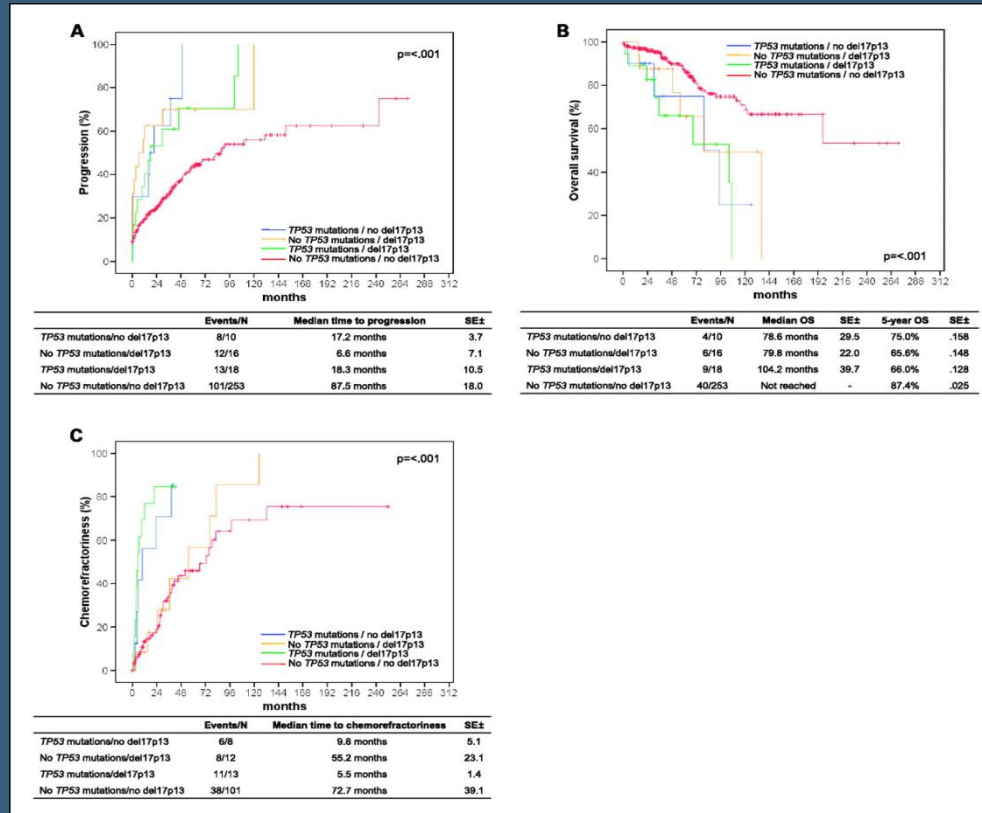
# 10 anni di EHA highlights: CLL

	Genetics					IF	Pre clinical/ Clinical studies
	TP53	telomeres	New genes	Stereotypy	Cytogenetics		
<b>2008</b>	As adverse as 17p-	Adverse prognosis		Prognosis (Richter)	t(14;19)	CD49d	Fluda+R Fluda+Thal
<b>2009-2010</b>	TP53 (trials)		prognosis in RS				FCR –
<b>2011</b>			WES / CE BRAF				Mo Abs R/R CAL-PIK3d BTK
<b>2012</b>			Cell of origin Prognosis				Genetic-driven treatment Chemoimmuno Rel/ref / BCL2
<b>2013</b>			Integrated classification				NOTCH1/CD20 CLL11 BCL2
<b>2014</b>	True predictor		mechanisms				Idela+R // Ibru vs ofa // Chlor + ofa ABT-199
<b>2015</b>	Small clones						FCR-IGHV mut
<b>2016</b>	CLL-IPI			prognosis			Acalabrutinib Obinutuzumab (green) MRD-OS / Venetoclax after BCR inhibitors
<b>2017</b>					Complex karyotype		FC obinu+ibru in IGHV mut // CLL-BAG Venetoclax MRD // Ibrutinib 4 y FU Ibru+venetoclax

# Genetics/ cytogenetics

10

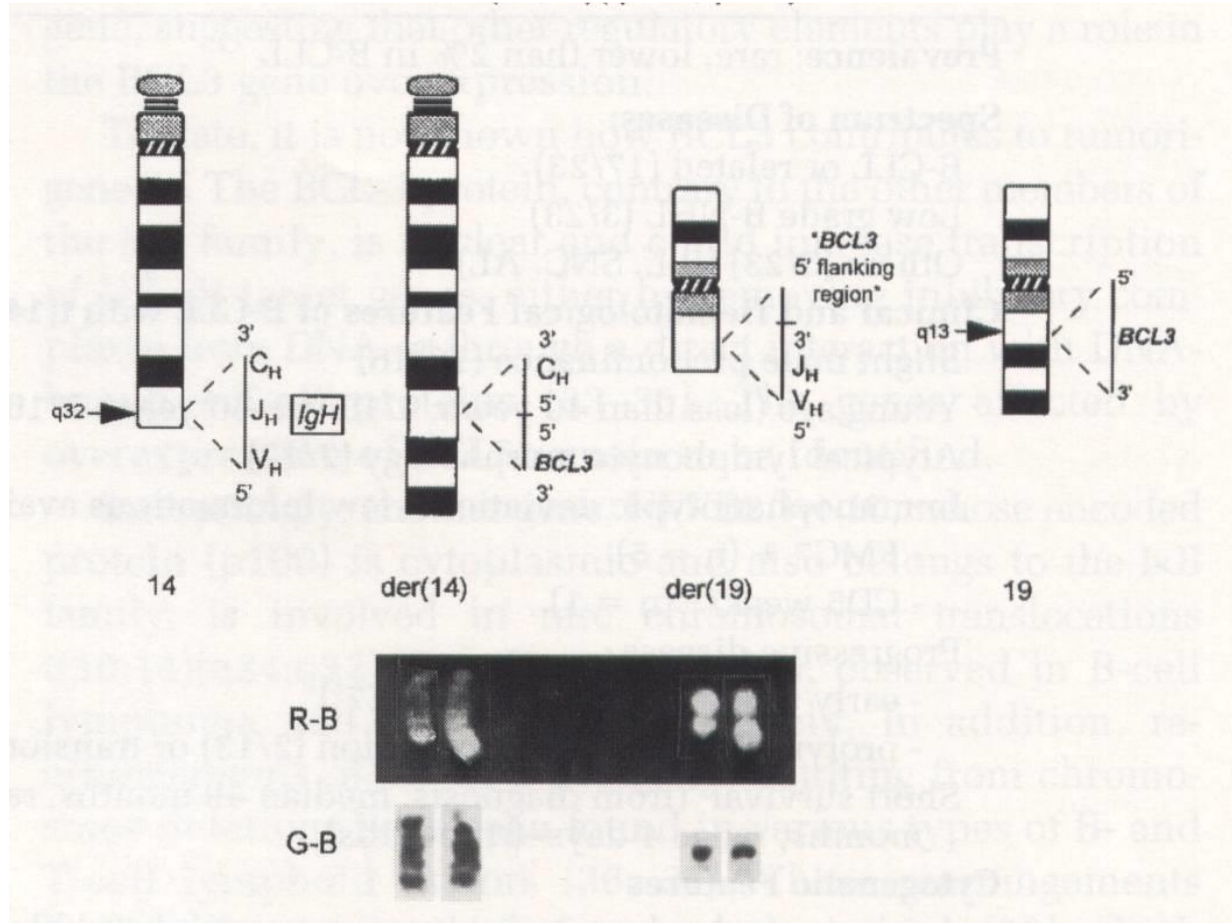
- >80% del17 carry TP53 mutations
- TP53 mutations as sole mutation in 4.5% CLL



- TP53 mutations are an independent prognostic factor

(Ghia P, adapted from Cerri et al EHA, 2008)

# t(14;19)(q32;q13)/*BCL3* in CLL



## **t(14;19)(q32;q13)/*BCL3* in CLL**

**Present in 0,1-2% of CLL**

**Rare as single; additional trisomy 12 in 50% of the cases**

**Overexpression of *BCL3* (modulation of NF-kB TF)**

**Atypical morphology**

**FMC7 positive – CD5 weak positivity**

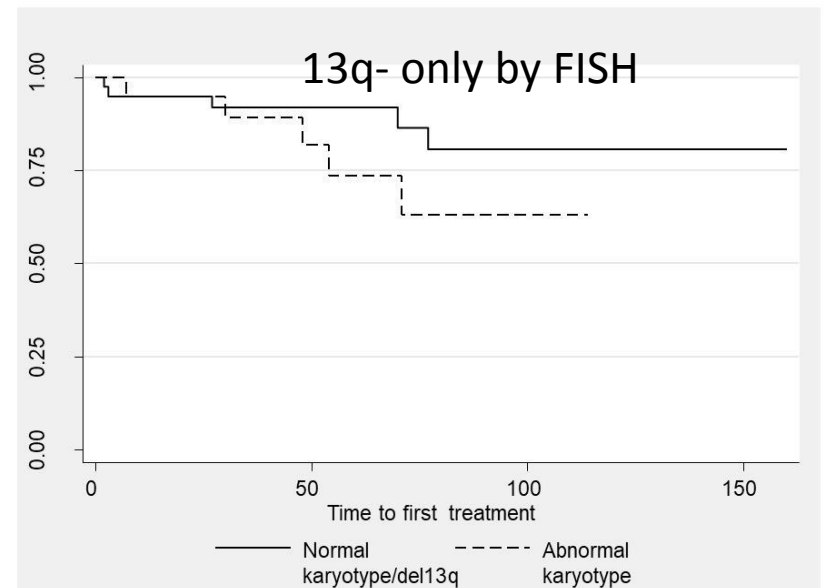
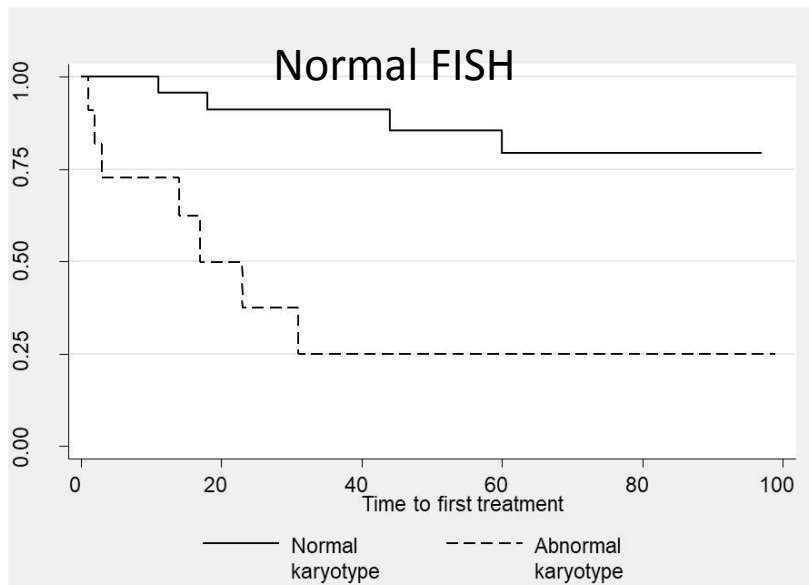
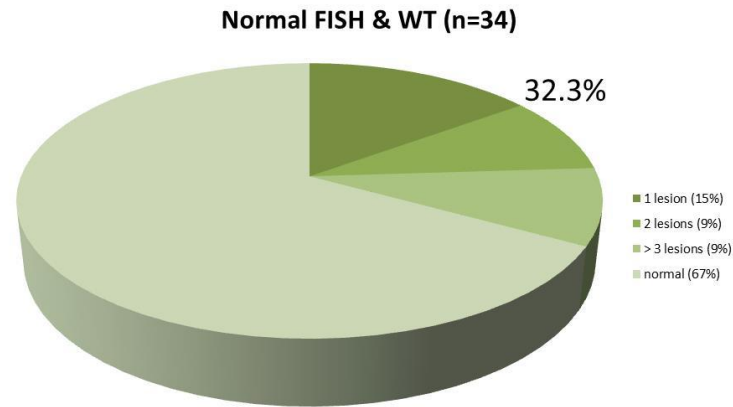
**Frequent in the young age**

**Cytologic transformation**

**Disease progression and short survival**

# INTEGRATION OF NOVEL GENE MUTATIONS INTO KARYOTYPE-BASED SUBGROUPS AS A PROGNOSTIC RISK STRATIFICATION TOOL IN TREATMENT-NAÏVE CLL

N = 153	n	%
17p-	5	3.3%
11q-	13	8.5%
+12	19	12.4%
Normal	46	30%
13q-	70	45.8%

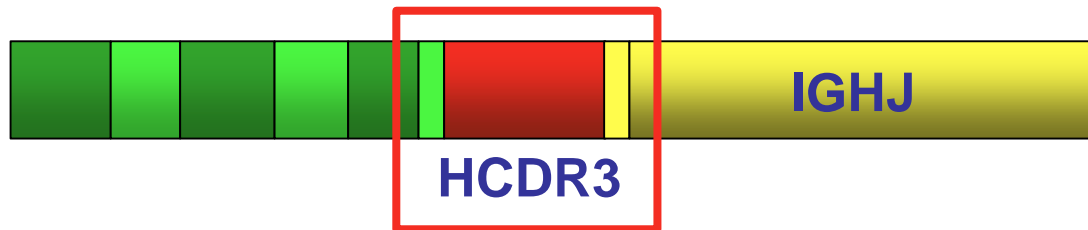


# “Stereotypy” in BCR Structure

Chance that two different B-cell clones might randomly use identical B cell receptors:  $10^{-10}$ - $10^{-12}$  (0.000000000001%)

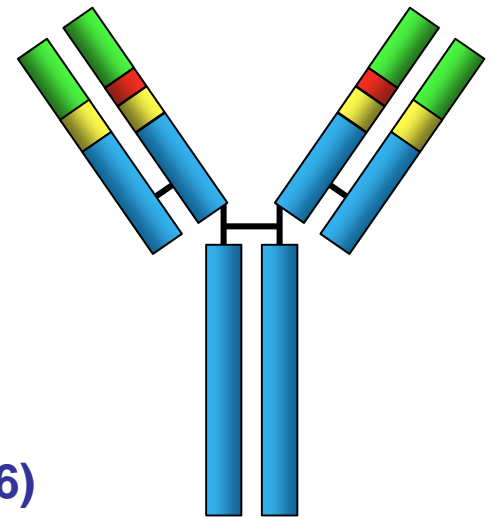
In CLL, this can occur once in every in 100-1000 clones

*Homologous complementarity determining region*



30.4% OF ALL CLL CASES (2308/7596)

~50% of U-CLL cases



# Stereotyped Receptors

14

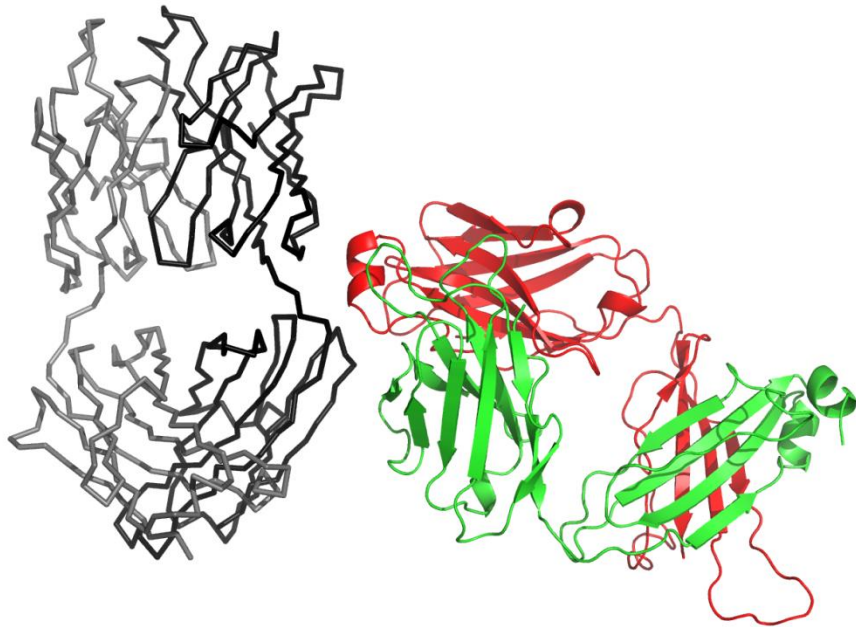
- Common herpesviruses and CLL: molecular evidence for a potential link with a subset of patients expressing stereotyped IGHV4-34 B cell receptors (Kostareli E., et al – Thessaloniki)
- CLL transforming to Richter's syndrome carry stereotyped HCDR3s at very high frequency (>50%) and display biased use of IGHV4-39 genes (Valeria S. et al – Novara)
- Novel molecular and clinical features of CLL expressing or not expressing stereotyped B cell receptors: results of an Italian multicentric study (Bomben R., et al – Aviano)
- Analysis of Chronic Lymphocytic Leukemias cases with stereotypic immunoglobulin's receptors in Ukrainian cohort (Bilous N., et al – Kiev)



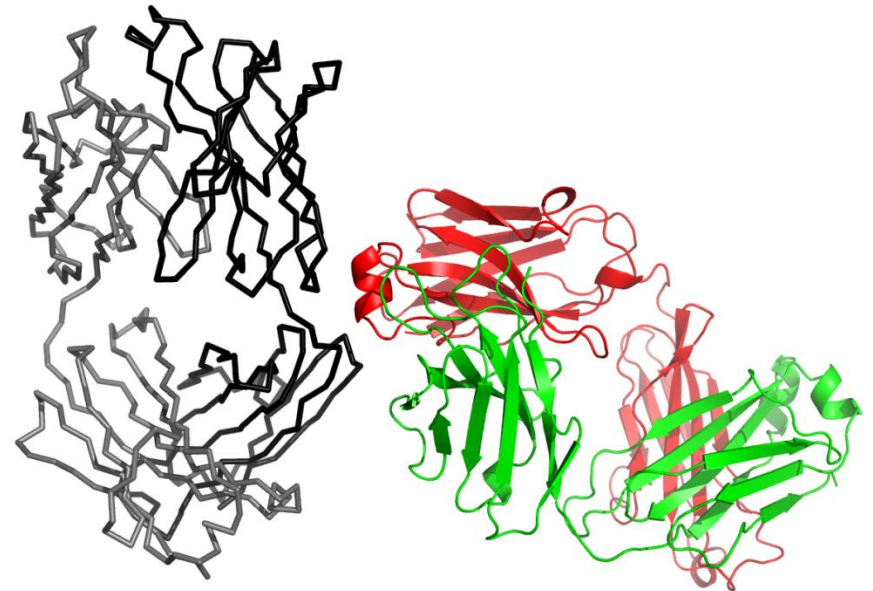
# Subset 4: self recognition of CLL Fab

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**CLL240**



**CLL183**



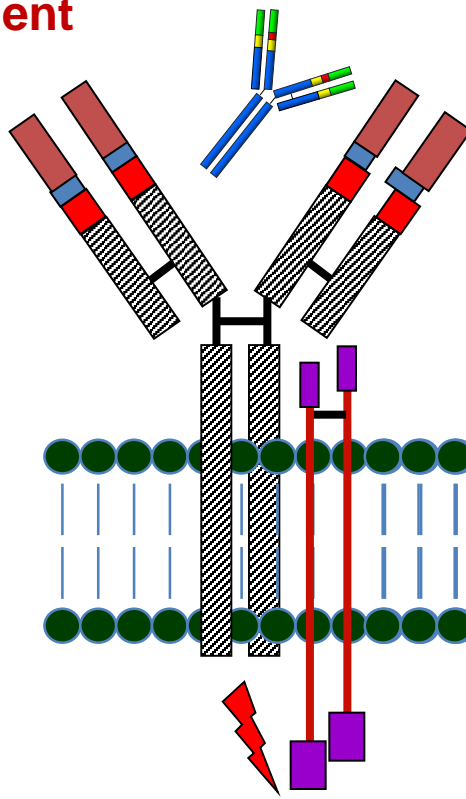
**Interaction with the V-C hinge (VH FR1 and CH1 domains)**

# BCR signalling in CLL is heterogeneous

**Aggressive**

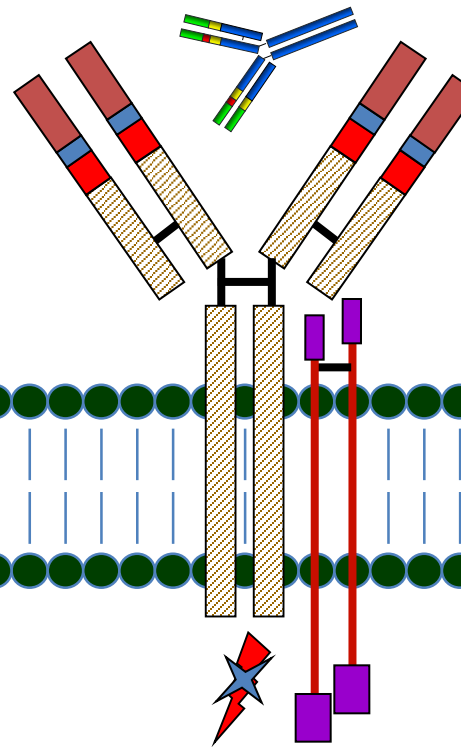
**Indolent (anergic)**

**weak, transient  
binding**



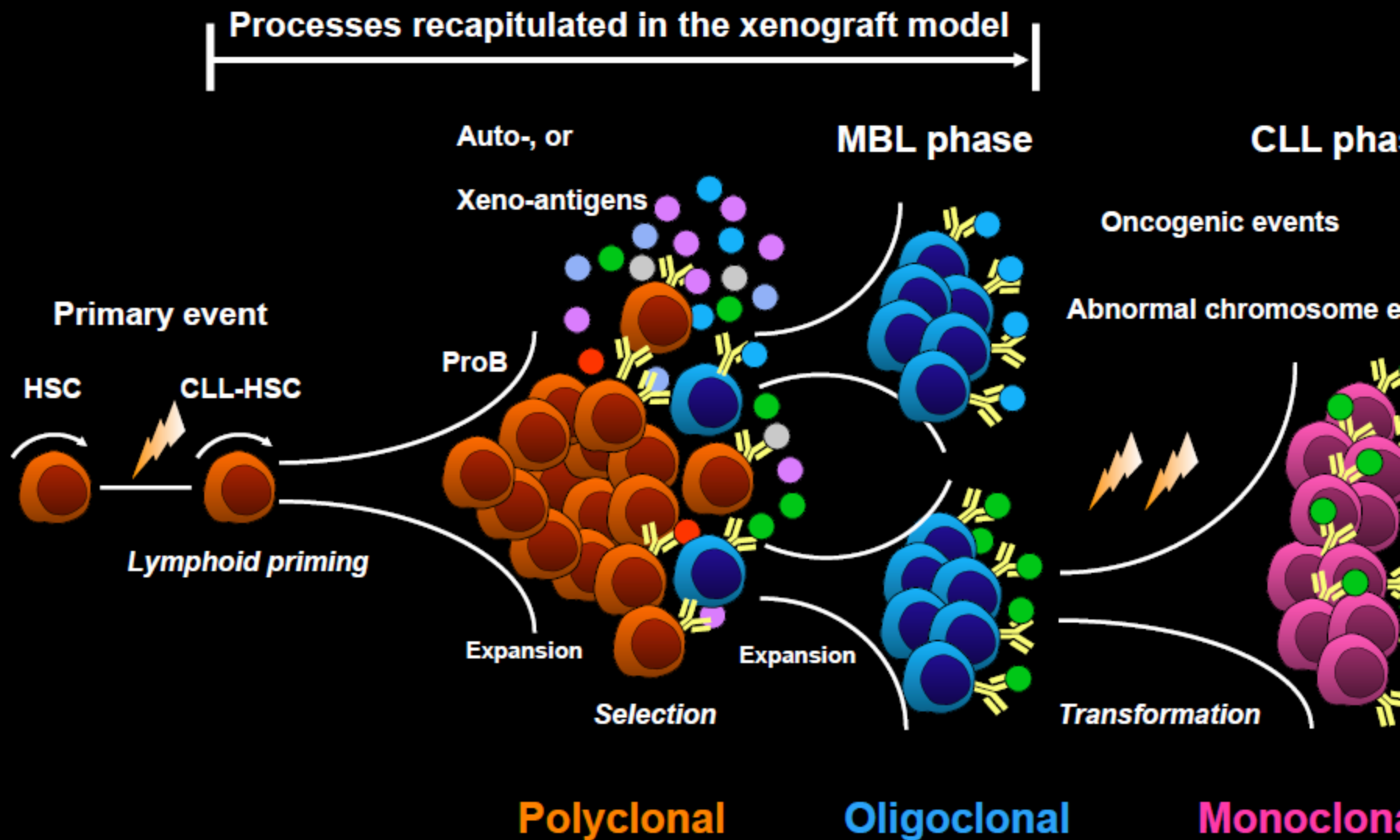
**Survival  
Proliferation**

**tight, stable  
binding**



Courtesy of Paolo Ghia

# Self-Renewing HSP is the Primary Target in the Pathogenesis of Human CLL



## Whole-genome sequencing identifies recurrent mutations in chronic lymphocytic leukaemia

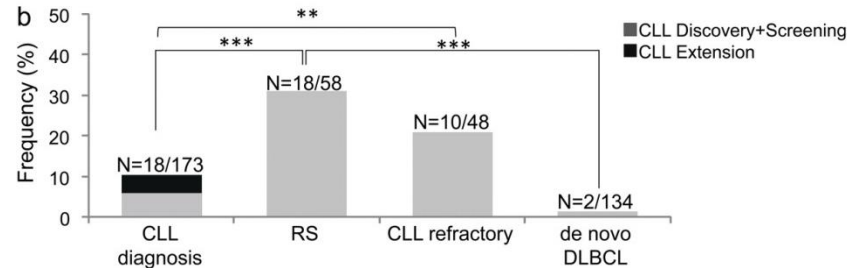
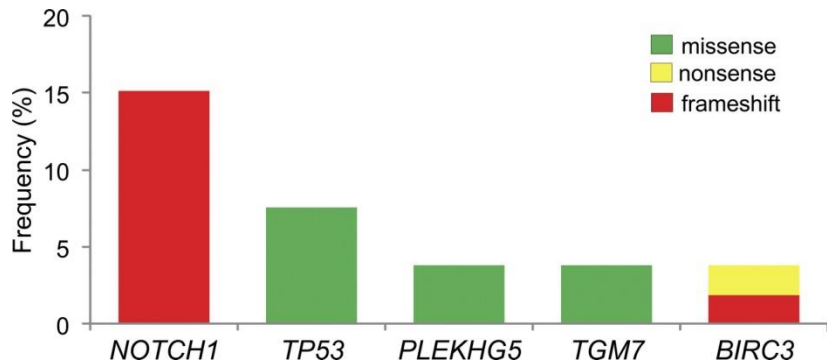
Xose S. Puente<sup>1</sup>, Magda Pinyol<sup>2</sup>, Víctor Quesada<sup>1</sup>, Laura Conde<sup>3</sup>, Gonzalo R. Ordóñez<sup>1</sup>, Neus Villamor<sup>3</sup>, Georgia Escaramis<sup>4</sup>, Pedro Jares<sup>3</sup>, Silvia Beà<sup>3</sup>, Marcos González-Díaz<sup>5</sup>, Laia Bassaganyas<sup>4</sup>, Tycho Baumann<sup>6</sup>, Manel Juan<sup>7</sup>, Mónica López-Guerra<sup>3</sup>, Dolors Colomer<sup>3</sup>, José M. C. Tubío<sup>4,8</sup>, Cristina López<sup>3</sup>, Alba Navarro<sup>3</sup>, Cristian Tornador<sup>4</sup>, Marta Aymerich<sup>3</sup>, María Rozman<sup>3</sup>, Jesús M. Hernández<sup>5</sup>, Diana A. Puente<sup>1</sup>, José M. P. Freije<sup>1</sup>, Gloria Velasco<sup>1</sup>, Ana Gutiérrez-Fernández<sup>1</sup>, Dolors Costa<sup>3</sup>, Anna Carrió<sup>3</sup>, Sara Guijarro<sup>3</sup>, Anna Enjuanes<sup>3</sup>, Lluís Hernández<sup>3</sup>, Jordi Yagüe<sup>7</sup>, Pilar Nicolás<sup>9</sup>, Carlos M. Romeo-Casabona<sup>9</sup>, Heinz Himmelbauer<sup>10</sup>, Ester Castillo<sup>10</sup>, Juliane C. Dohm<sup>10</sup>, Silvia de Sanjosé<sup>11</sup>, Miguel A. Piris<sup>12</sup>, Enrique de Alava<sup>5</sup>, Jesús San Miguel<sup>5</sup>, Romina Royo<sup>13</sup>, Josep L. Gelpi<sup>13</sup>, David Torrents<sup>13</sup>, Modesto Orozco<sup>13</sup>, David G. Pisano<sup>14</sup>, Alfonso Valencia<sup>14</sup>, Roderic Guigó<sup>15</sup>, Mónica Bayés<sup>16</sup>, Simon Heath<sup>16</sup>, Marta Gut<sup>16</sup>, Peter Klatt<sup>17</sup>, John Marshall<sup>18</sup>, Keiran Raine<sup>18</sup>, Lucy A. Stebbings<sup>18</sup>, P. Andrew Futreal<sup>18</sup>, Michael R. Stratton<sup>18</sup>, Peter J. Campbell<sup>18</sup>, Ivo Gut<sup>16</sup>, Armando López-Guillermo<sup>6</sup>, Xavier Estivill<sup>4</sup>, Emili Montserrat<sup>6</sup>, Carlos López-Otín<sup>1\*</sup> & Elías Campo<sup>3\*</sup>

**Table 1 | Genes recurrently mutated in chronic lymphocytic leukaemia**

Gene	Protein	Mutation	Mutated cases / total	Overall frequency (%)	Frequency in IGHV-unmutated (%)	Frequency in IGHV-mutated (%)
<i>NOTCH1</i>	Notch 1	P2515Rfs*4 Q2503*	29/255 1/255	12.2	20.4	7
<i>MYD88</i>	Myeloid differentiation primary response gene 88	F2482Ffs*2 L265P	1/255 9/310	2.9	0.8	5.6
<i>XPO1</i>	Exportin 1	E571K E571G	3/165 1/165	2.4	4.6	0
<i>KLHL6</i>	Kelch-like 6	F49L/L65P L90F L58P/T64A/Q81P	3/160	1.8	0	4.5

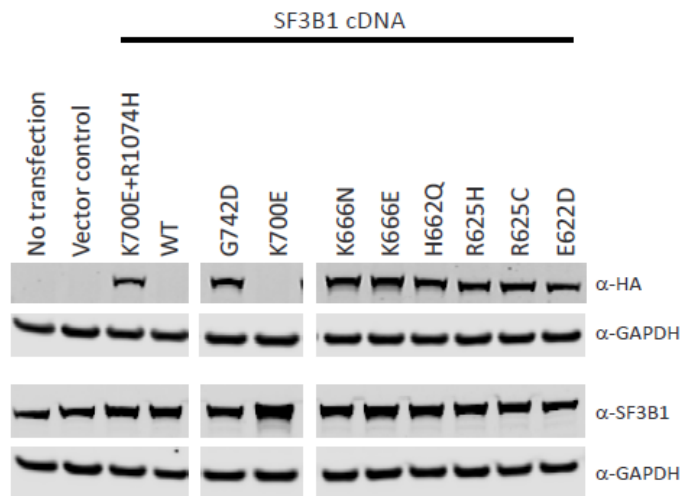
# Analysis of the chronic lymphocytic leukemia coding genome: role of *NOTCH1* mutational activation

Giulia Fabbri,<sup>1</sup> Silvia Rasi,<sup>5</sup> Davide Rossi,<sup>5</sup> Vladimir Trifonov,<sup>2</sup> Hossein Khiabani,<sup>2</sup> Jing Ma,<sup>6</sup> Adina Grunn,<sup>1</sup> Marco Fangazio,<sup>5</sup> Daniela Capello,<sup>5</sup> Sara Monti,<sup>5</sup> Stefania Cresta,<sup>5</sup> Ernesto Gargiulo,<sup>5</sup> Francesco Forconi,<sup>7</sup> Anna Guarini,<sup>8</sup> Luca Arcaini,<sup>9</sup> Marco Paulli,<sup>10</sup> Luca Laurenti,<sup>11</sup> Luigi M. Larocca,<sup>12</sup> Roberto Marasca,<sup>13</sup> Valter Gattei,<sup>14</sup> David Oscier,<sup>15</sup> Francesco Bertoni,<sup>16</sup> Charles G. Mullighan,<sup>6</sup> Robin Foà,<sup>8</sup> Laura Pasqualucci,<sup>1,3</sup> Raul Rabadan,<sup>2</sup> Riccardo Dalla-Favera,<sup>1,3,4</sup> and Gianluca Gaidano<sup>5</sup>

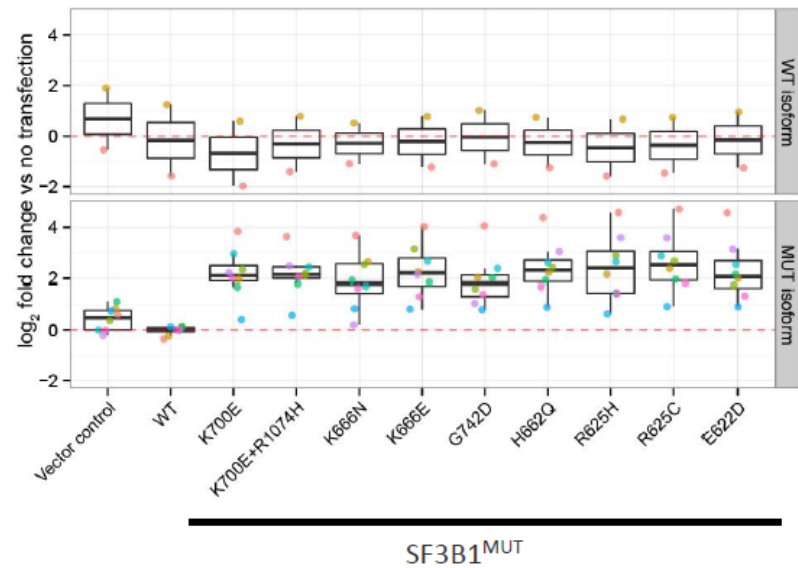


## Expression of SF3B1MUT upregulates mutant associated splice isoforms

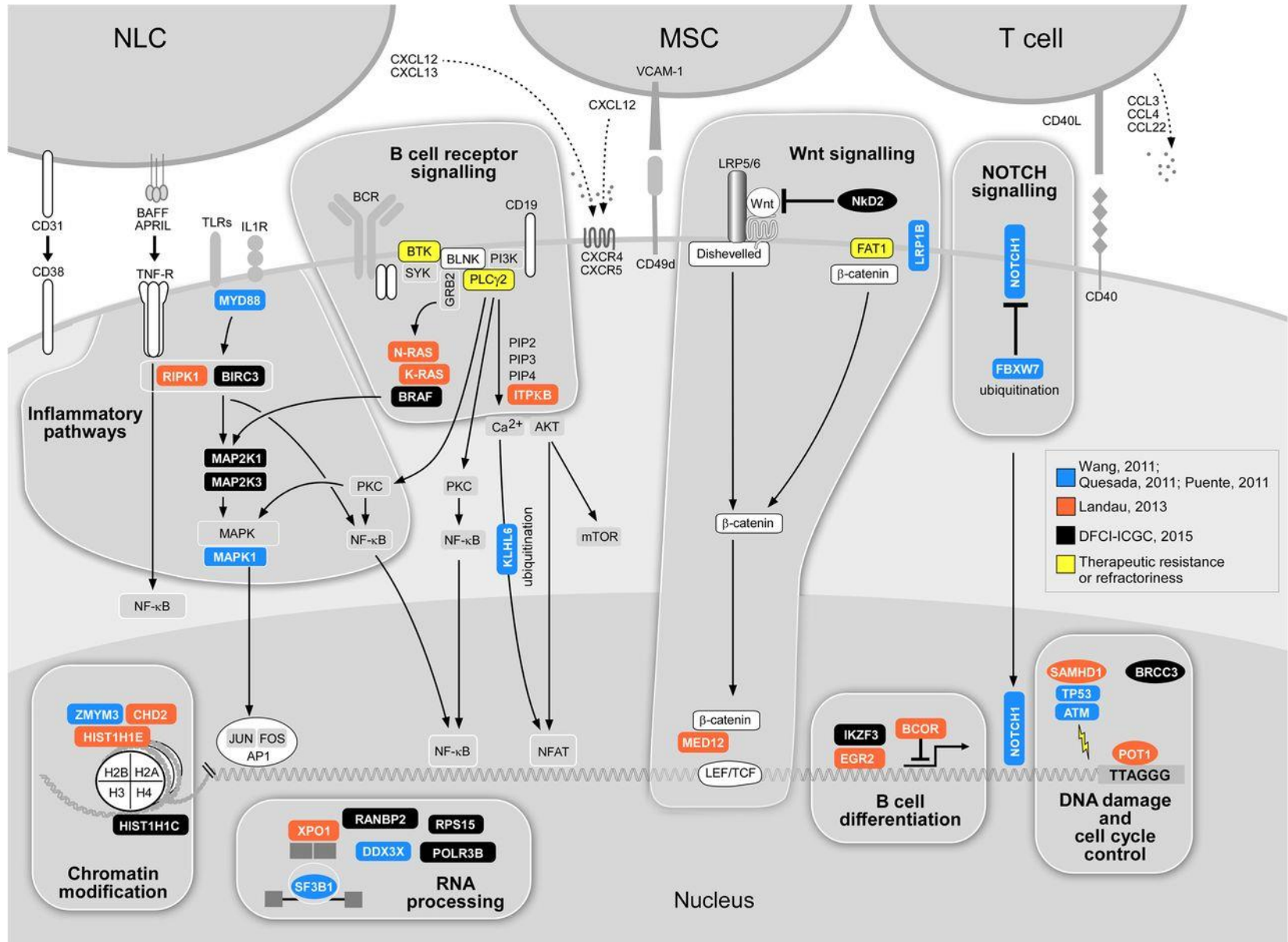
293FT cells transfected  
with SF3B1 cDNA  
48hr ↓  
Total RNA extracted for  
Nanostring



Expression of WT and MUT isoforms  
(Nanostring assay)

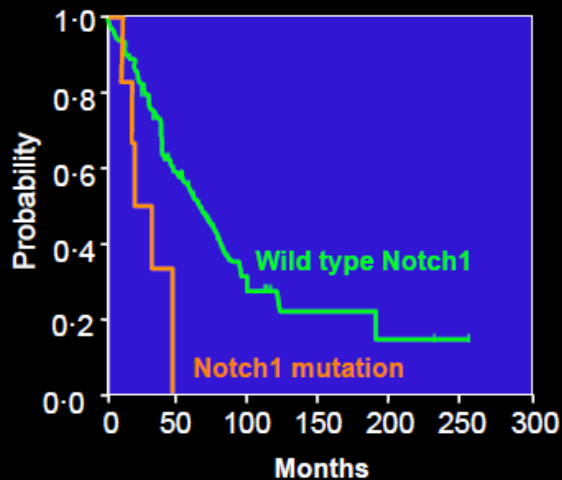


# Putative core cellular pathways affected by significantly mutated genes in CLL.

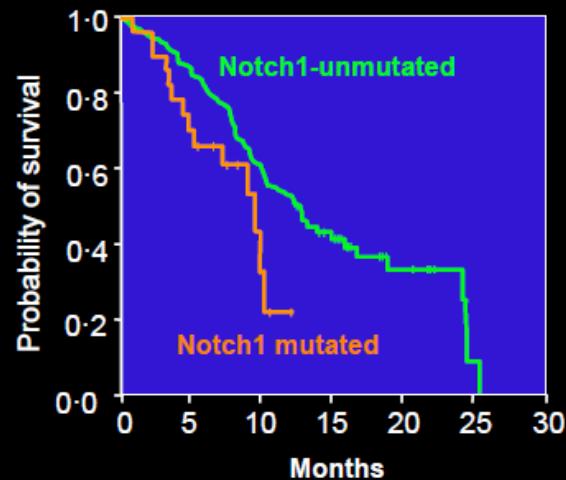


## Pivotal Studies Suggest a Prognostic Role of *NOTCH1* Mutations in CLL

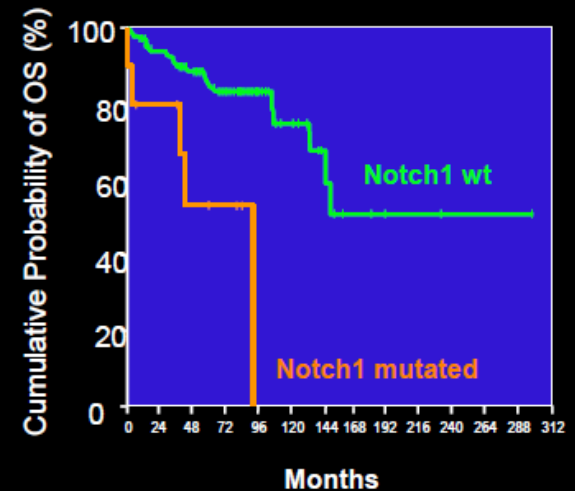
Sportoletti P. et al., Br J Haematol 2010



Puente XS. et al., Nature 2011



Fabbri G. et al., J Exp Med 2011



Study	N. patients	<i>NOTCH1</i> mutation	Clinical endpoint	Multivariate analysis
<i>Sportoletti P, 2010</i>	133	5%	TFS	No
<i>Puente XS, 2011</i>	255	12%	OS	No
<i>Fabbri G, 2011</i>	120	10%	TFS & OS	Yes

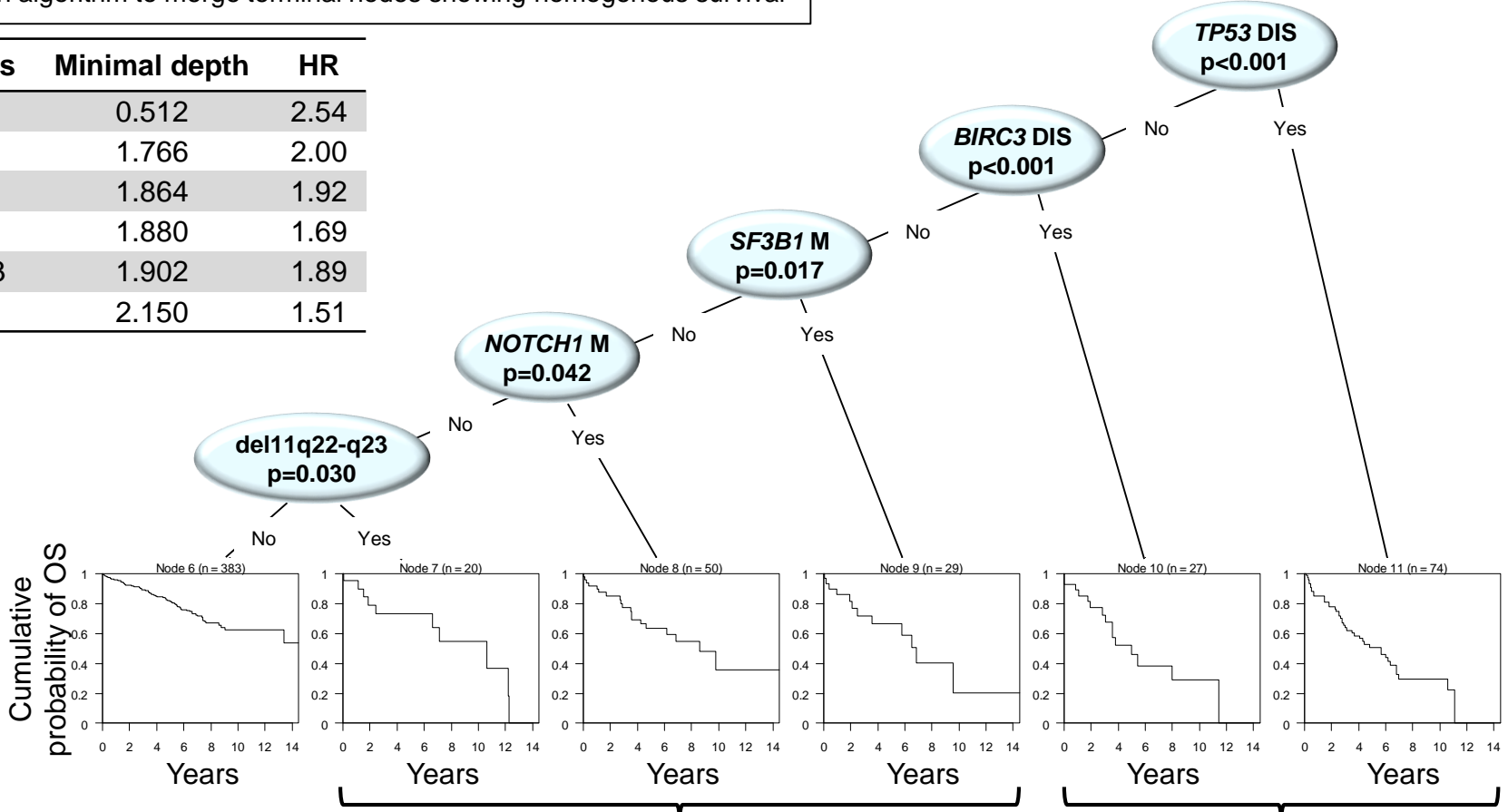




# Hierarchical order of relevance of genetic lesions in predicting survival in newly diagnosed CLL

- Recursive partitioning to divide patients into genetic subgroups with different OS
- Random survival forest validation of the stability of the recursive decision tree
- Amalgamation algorithm to merge terminal nodes showing homogenous survival

Top variables	Minimal depth	HR
<i>TP53</i> DIS	0.512	2.54
<i>BIRC3</i> DIS	1.766	2.00
<i>SF3B1</i> M	1.864	1.92
<i>NOTCH1</i> M	1.880	1.69
del11q22-q23	1.902	1.89
+12	2.150	1.51



***SF3B1* M  
and/or  
*NOTCH1* M  
and/or  
del11q22-q23**

***TP53* DIS  
and/or  
*BIRC3* DIS**

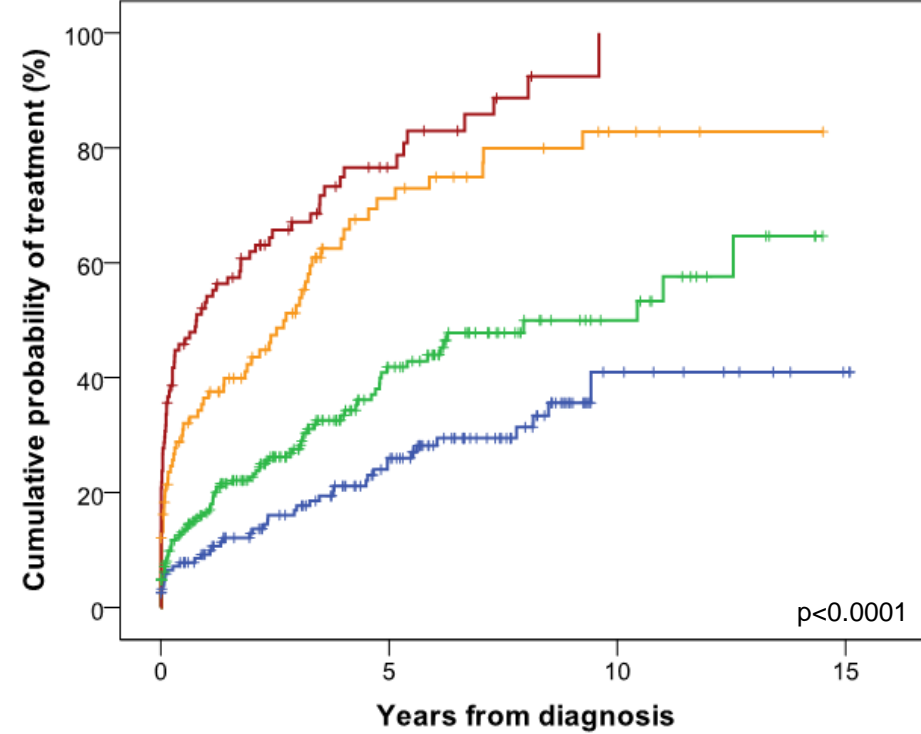
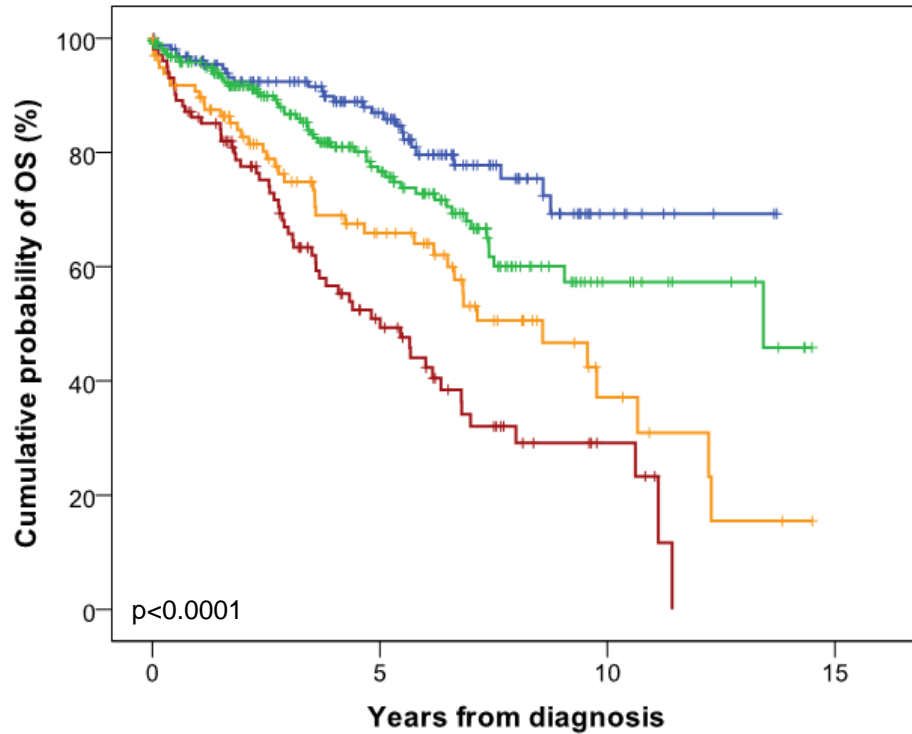
# Integrated mutational and cytogenetic model for CLL prognostication *(Rossi et al, Blood 2013)*

OS

Treatment

- del13q14
- Normal/+12
- NOTCH1 M/SF3B1 M/del11q22-q23
- TP53 DIS/BIRC3 DIS

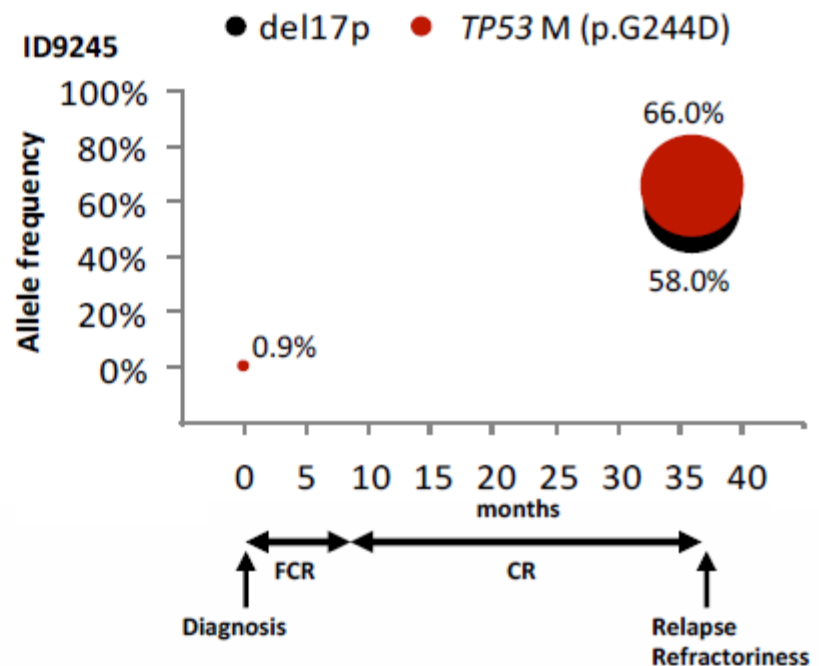
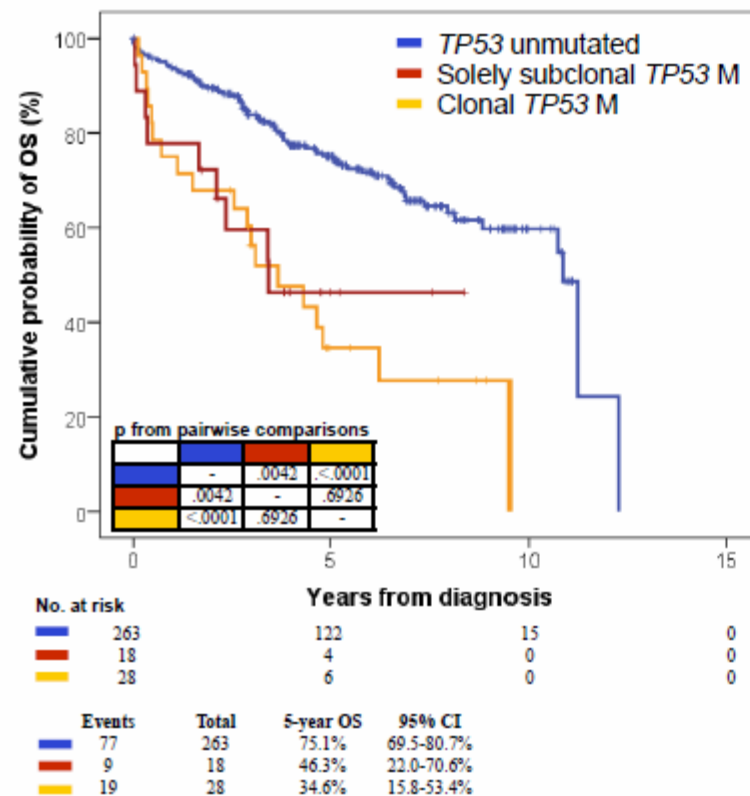
- del13q14
- Normal/+12
- NOTCH1 M/SF3B1 M/del11q22-q23
- TP53 DIS/BIRC3 DIS



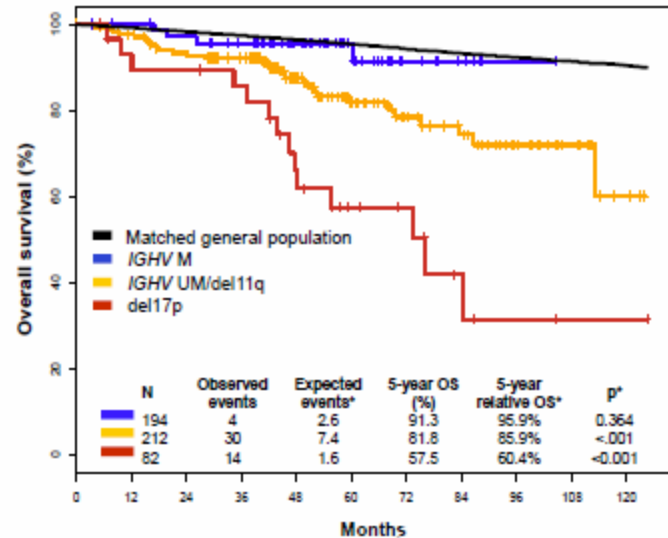
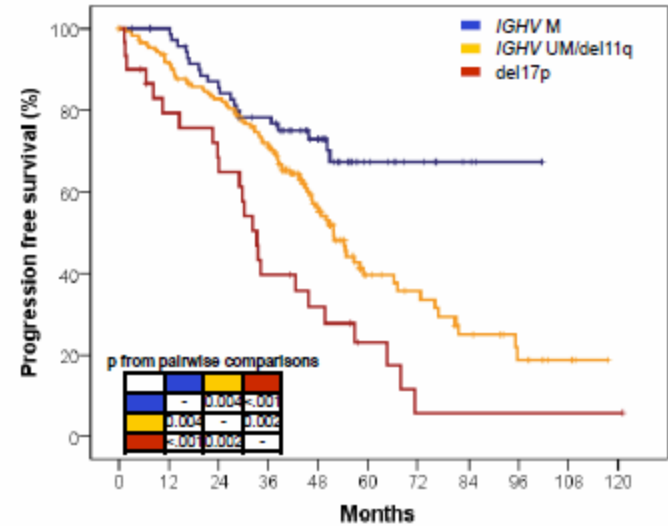
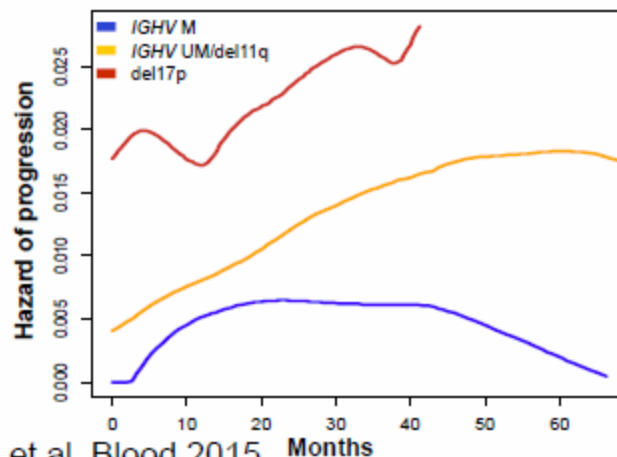
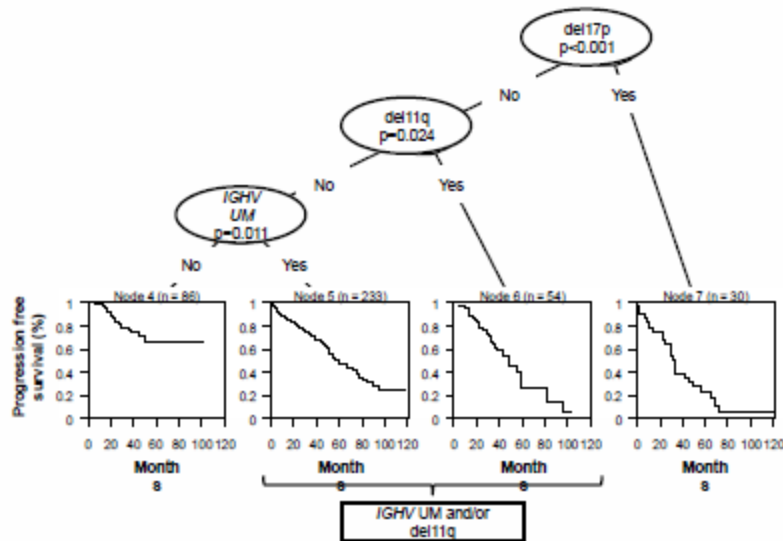
	N	%	10-years OS
del13q14	155	26%	69%
Normal/+12	228	39%	57%
NOTCH1 M/SF3B1 M/del11q22-q23	99	17%	37%
TP53 DIS/BIRC3 DIS	101	17%	29%

	N	%	Treated at 10 years
del13q14	155	26%	41%
Normal/+12	228	39%	50%
NOTCH1 M/SF3B1 M/del11q22-q23	99	17%	83%
TP53 DIS/BIRC3 DIS	101	17%	100%

## Small *TP53* mutated subclones have the same unfavorable prognostic impact as clonal *TP53* defects

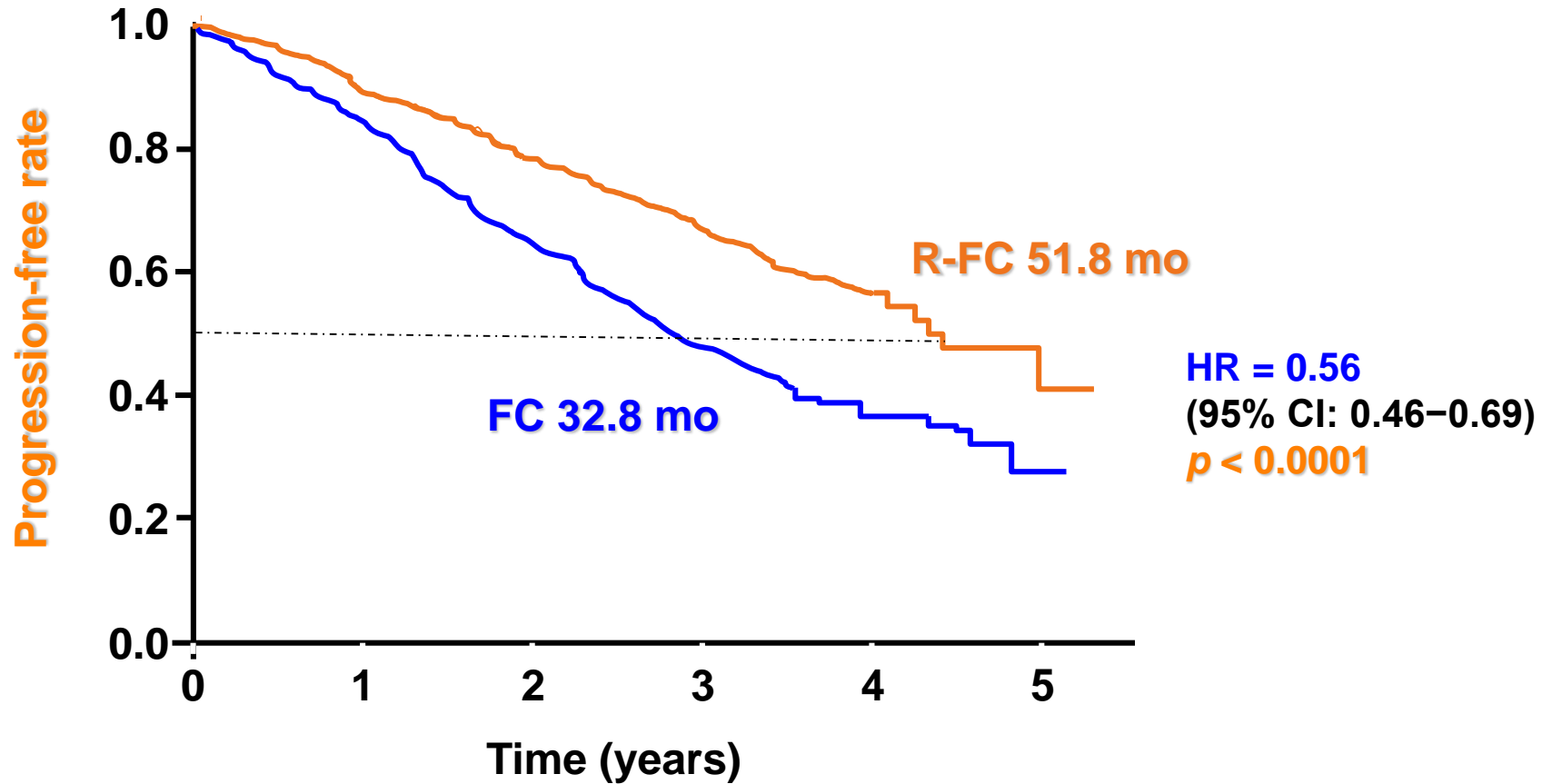


# IGHV mutated patients devoid of del17p and del11q gain the greatest benefit from FCR

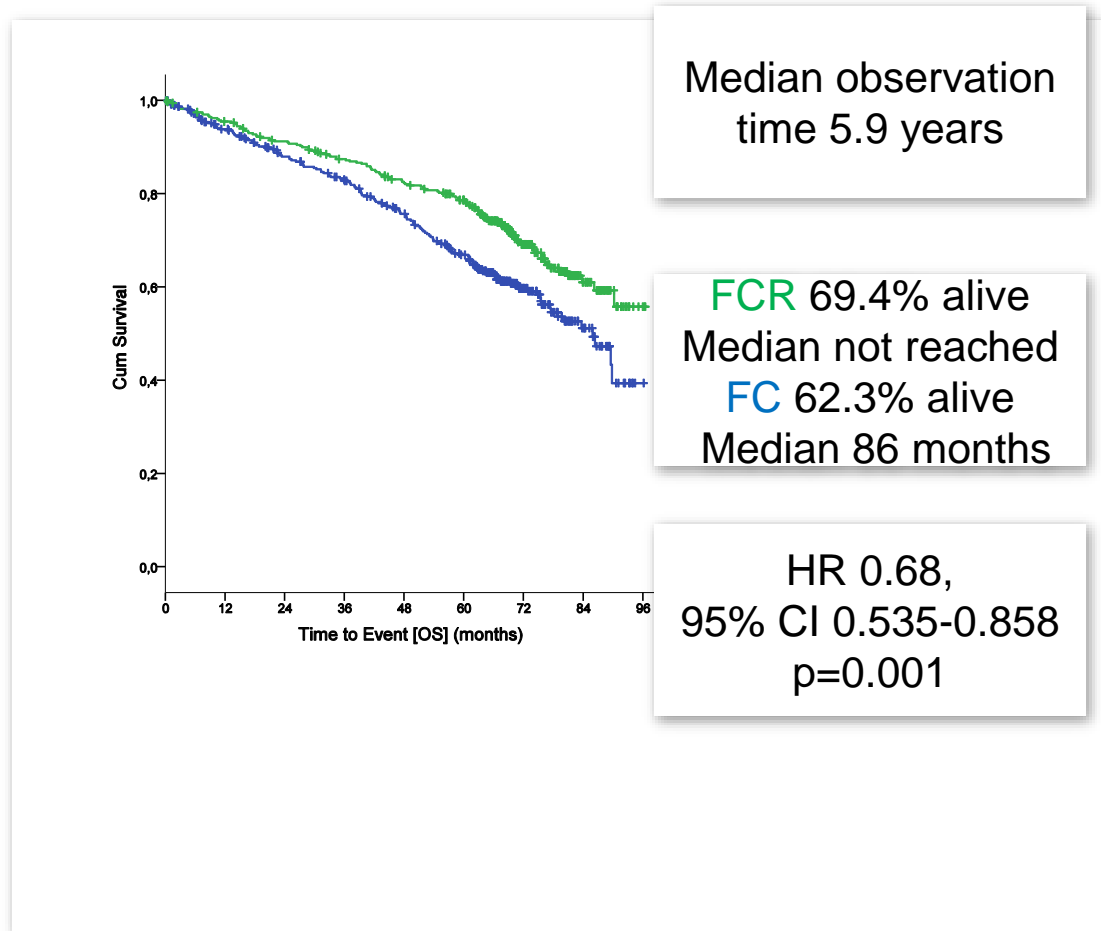


# CLL8: Progression-free Survival

*Median follow-up 3 years*



# CLL8 trial: Overall survival, update 2012



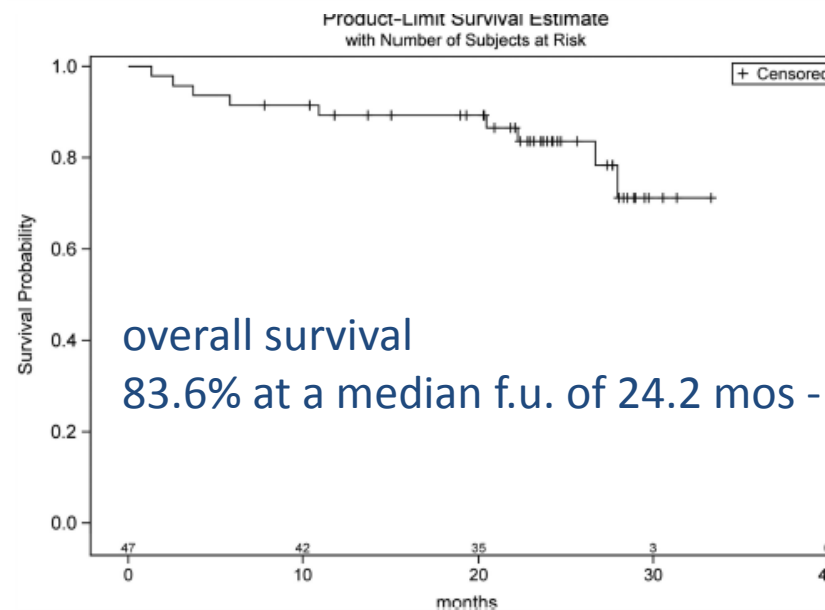
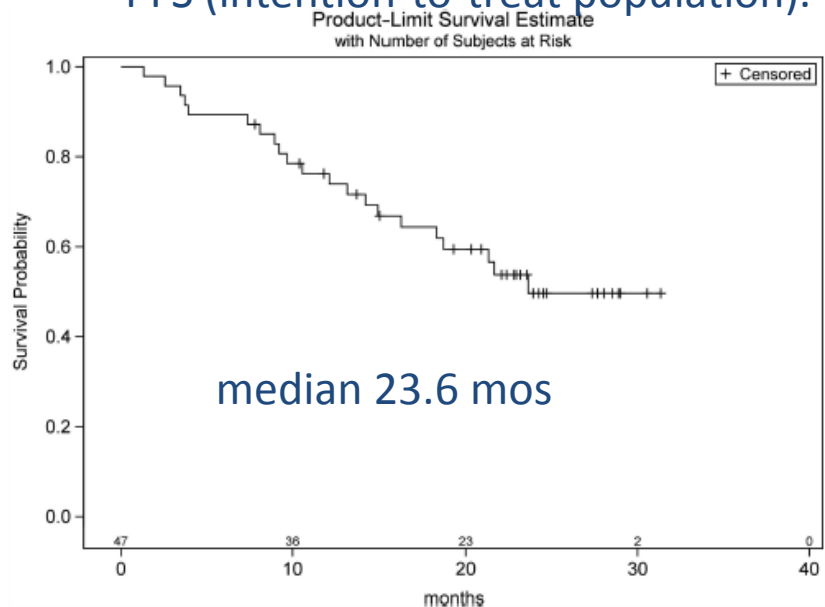
# A SINGLE-ARM MULTI-CENTER TRIAL OF BENDAMUSTINE GIVEN WITH OFATUMUMAB) IN PATIENTS WITH REFRACTORY OR RELAPSED CLL. GIMEMA CLL0809 PROTOCOL

**Table 1.** Patient demographic and baseline clinical characteristics

Characteristic	No. of patients	% Of patients
<b>Sex</b>		
Male	35	71
Female	14	29
<b>Age, years</b>		
Median (range)	66 (46-81)	35
≥70	17	65
<70	32	
<b>Previous treatment lines</b>		
1	30	61
2	19	39
<b>Previous chemotherapy</b>		
Fludarabine-based	37	75
Rituximab-based	27	55
Alemtuzumab-based	6	12
<b>IGHV mutational status</b>		
Mutated	17	35
Unmutated	32	65
Expression of ZAP-70 (>20%)	30	61
<b>Genomic aberrations by FISH</b>		
del(11q) <sup>23</sup>	6	12
del(13q)	23	47
del(17p)	9	18
+12	9	18
Expression of CD38 (>30%)	21	43
NOTCH1 mutations	8	16
BIRC3 mutations	3	6
SF3B1 mutations	11	22
TP53 mutations	9	18

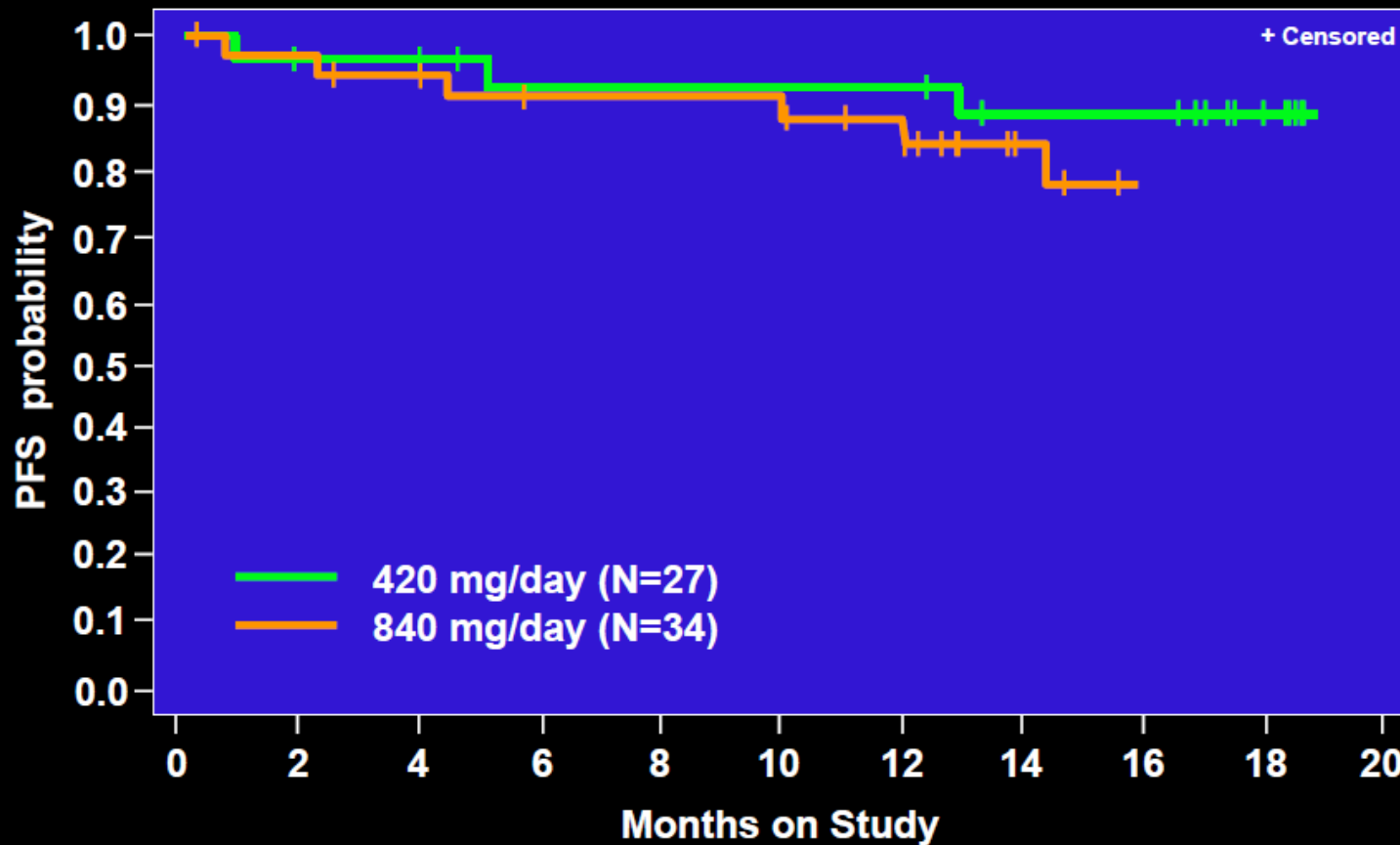
## PFS (intention-to-treat population).

median follow-up of 24.2 months



# PCYC-1102-CA: Progression-Free Survival (Relapsed/Refractory) by Dose

*Estimated 18 mo PFS at 420 mg/d = 87.7%*



S. O'Brien, EHA Amsterdam, June 14-17 (2012)

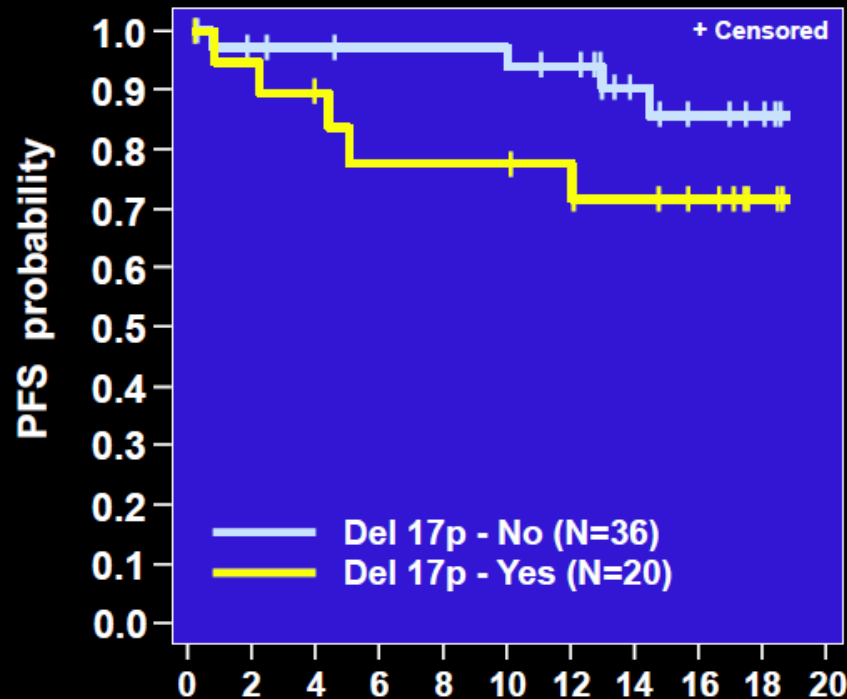


Università di Padova

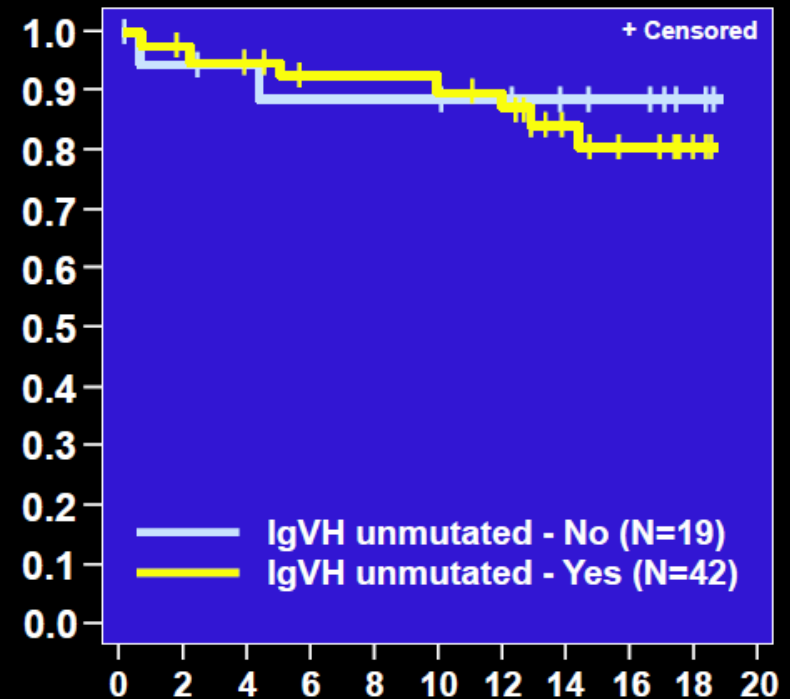


# PCYC-1102-CA: Progression-Free Survival (Relapsed/Refractory) by Risk Factors

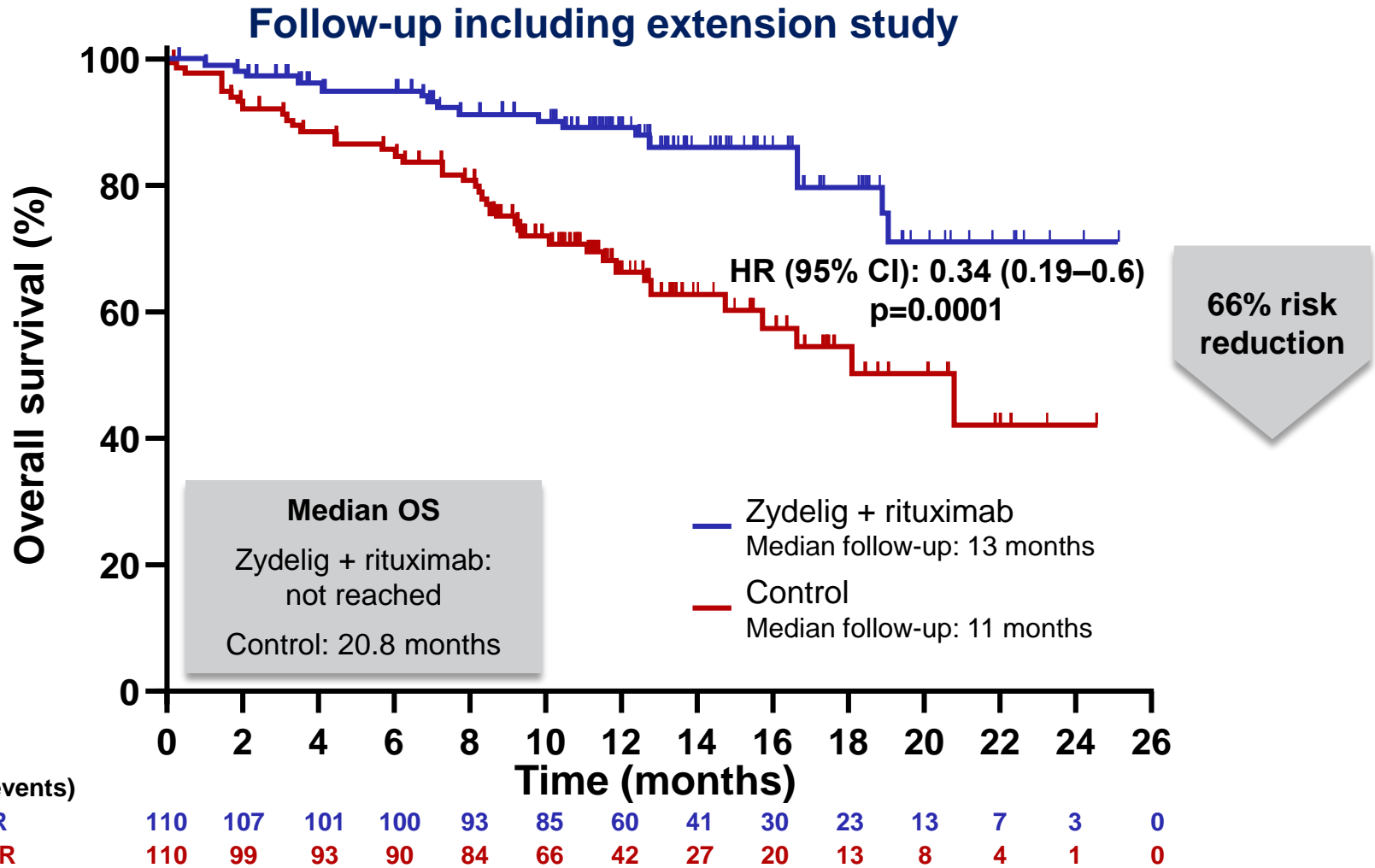
## Del 17p Status



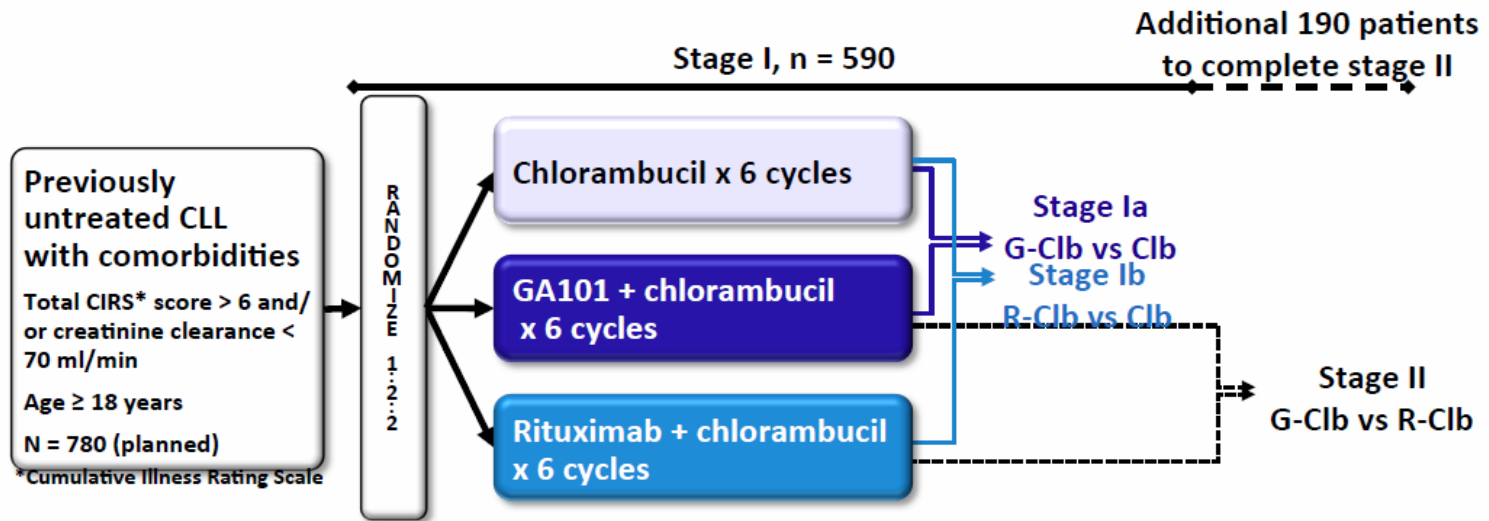
## IgVH Status



# Difference in efficacy of Zydelig + rituximab maintained despite crossover in the extension study

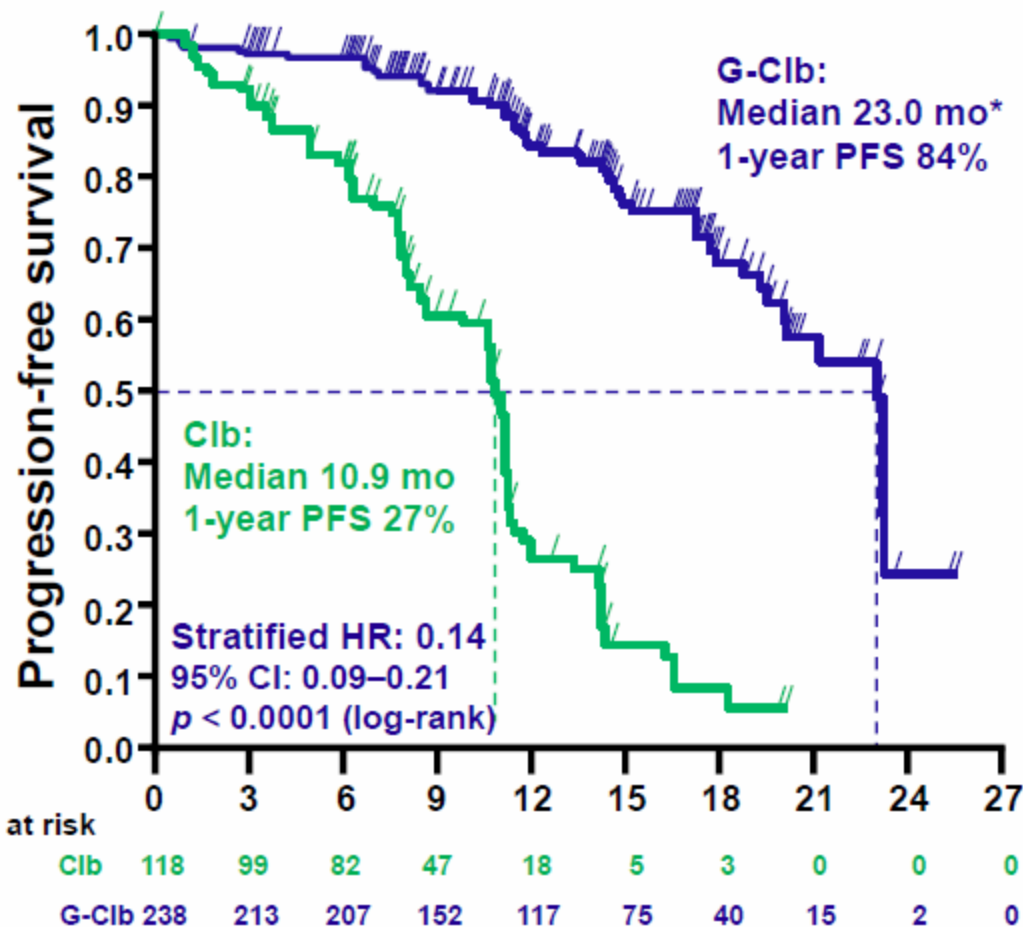


## CLL11: Study design

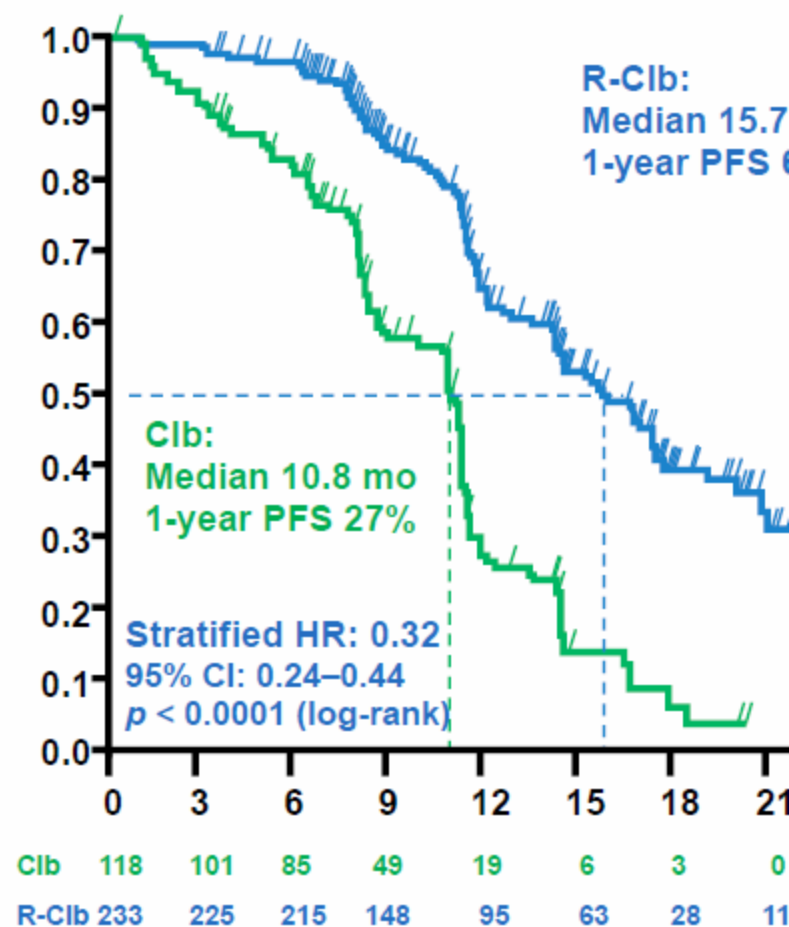


- GA101: 1,000 mg days 1, 8, and 15 cycle 1; day 1 cycles 2–6, every 28 days
- Rituximab: 375 mg/m<sup>2</sup> day 1 cycle 1, 500 mg/m<sup>2</sup> day 1 cycles 2–6, every 28 days
- Clb: 0.5 mg/kg day 1 and day 15 cycle 1–6, every 28 days
- Patients with progressive disease in the Clb arm were allowed to cross over to G-Clb

## Stage Ia



## Stage Ib

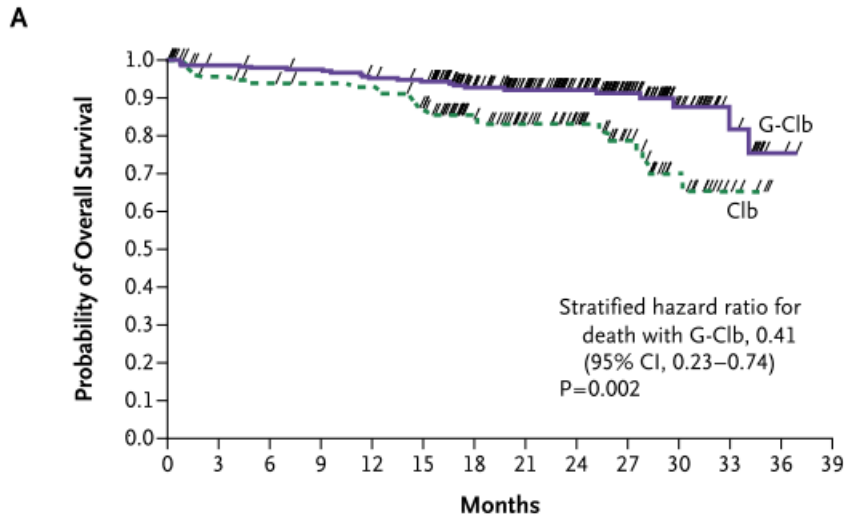


Type 1 error controlled through closed test procedure;  $p$ -value of the global test was  $<.0001$ .

\* In the G-Clb arm  $< 10\%$  of patients had reached the median at cutoff; therefore, in contrast to the Clb arm the G-Clb median PFS could not be reliably estimated due to the few patients at risk at time of G-Clb median.

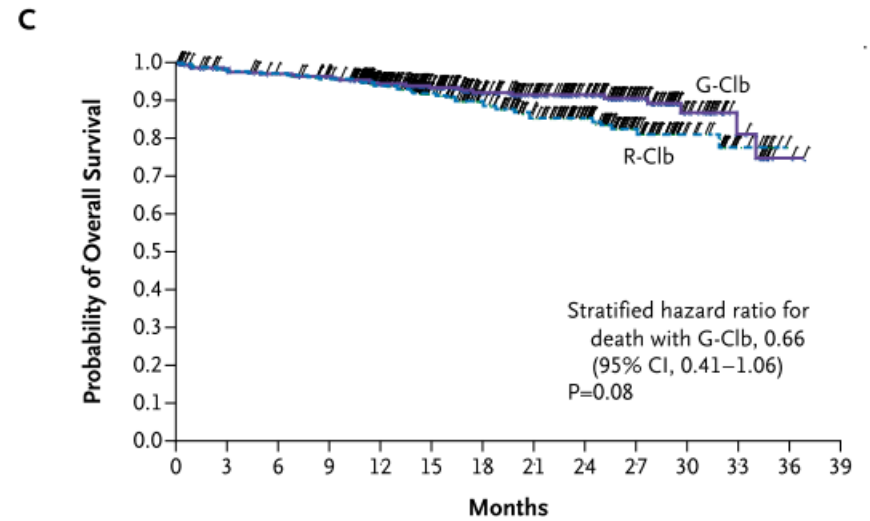
# Improved survival using Obinutuzumab plus Chlorambucil in Patients with CLL and Coexisting Conditions

The NEW ENGLAND JOURNAL of MEDICINE



No. at Risk

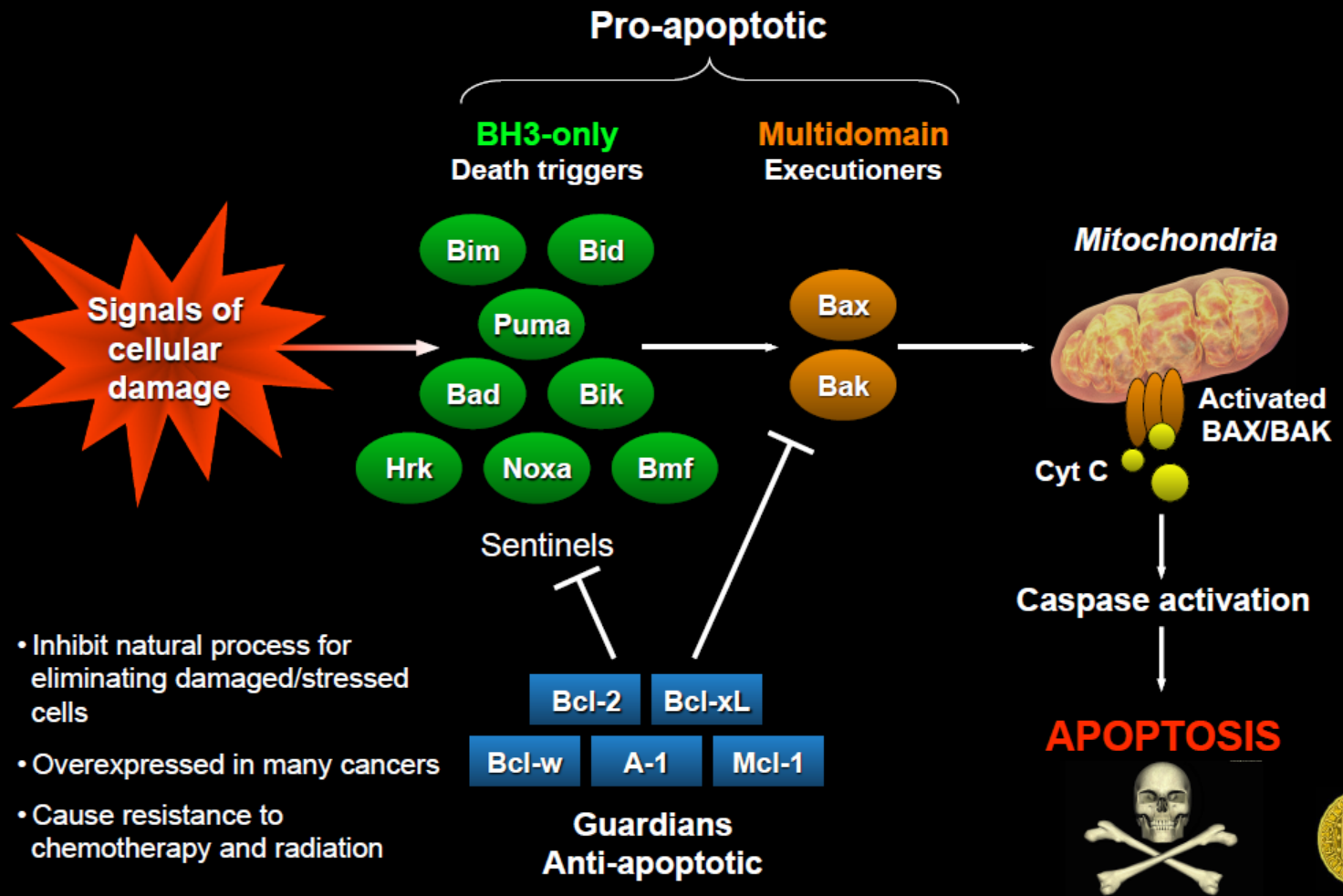
G-Clb	238	226	223	221	215	211	170	144	115	71	34	14	2	0
Clb	118	109	105	103	102	94	70	56	44	29	15	5	0	0



No. at Risk

G-Clb	333	316	310	303	261	214	170	144	115	71	34	14	2	0
R-Clb	330	320	314	305	255	203	169	138	105	61	27	8	0	0

# Bcl-2 family proteins regulate apoptosis



- Inhibit natural process for eliminating damaged/stressed cells
- Overexpressed in many cancers
- Cause resistance to chemotherapy and radiation



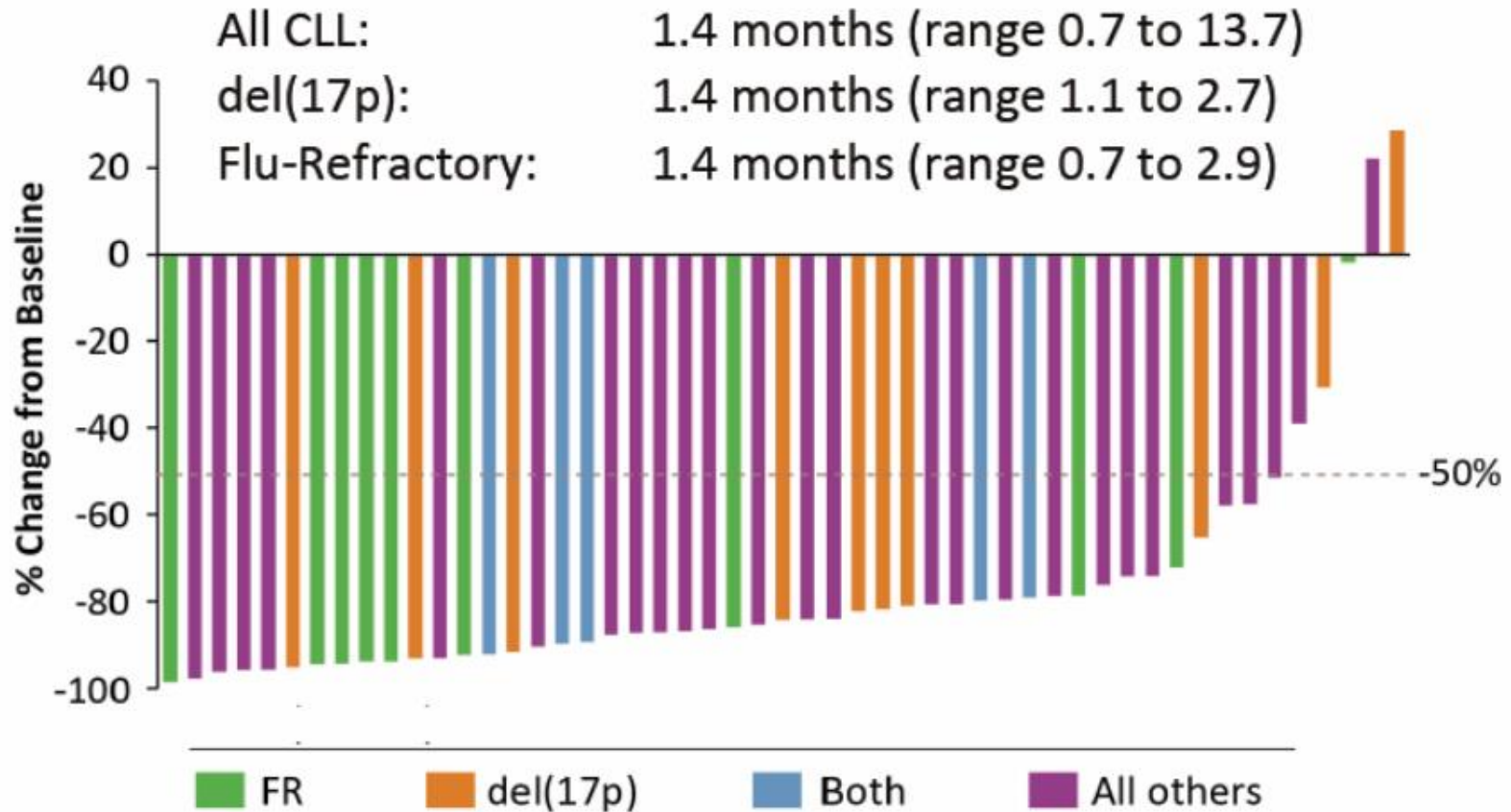
el

The BCL-2 inhibitor ABT-199 (GDC-0199) is active and well tolerated in ultra high risk relapsed/refractory chronic lymphocytic leukemia (CLL) (Roberts, Melbourne)

2013

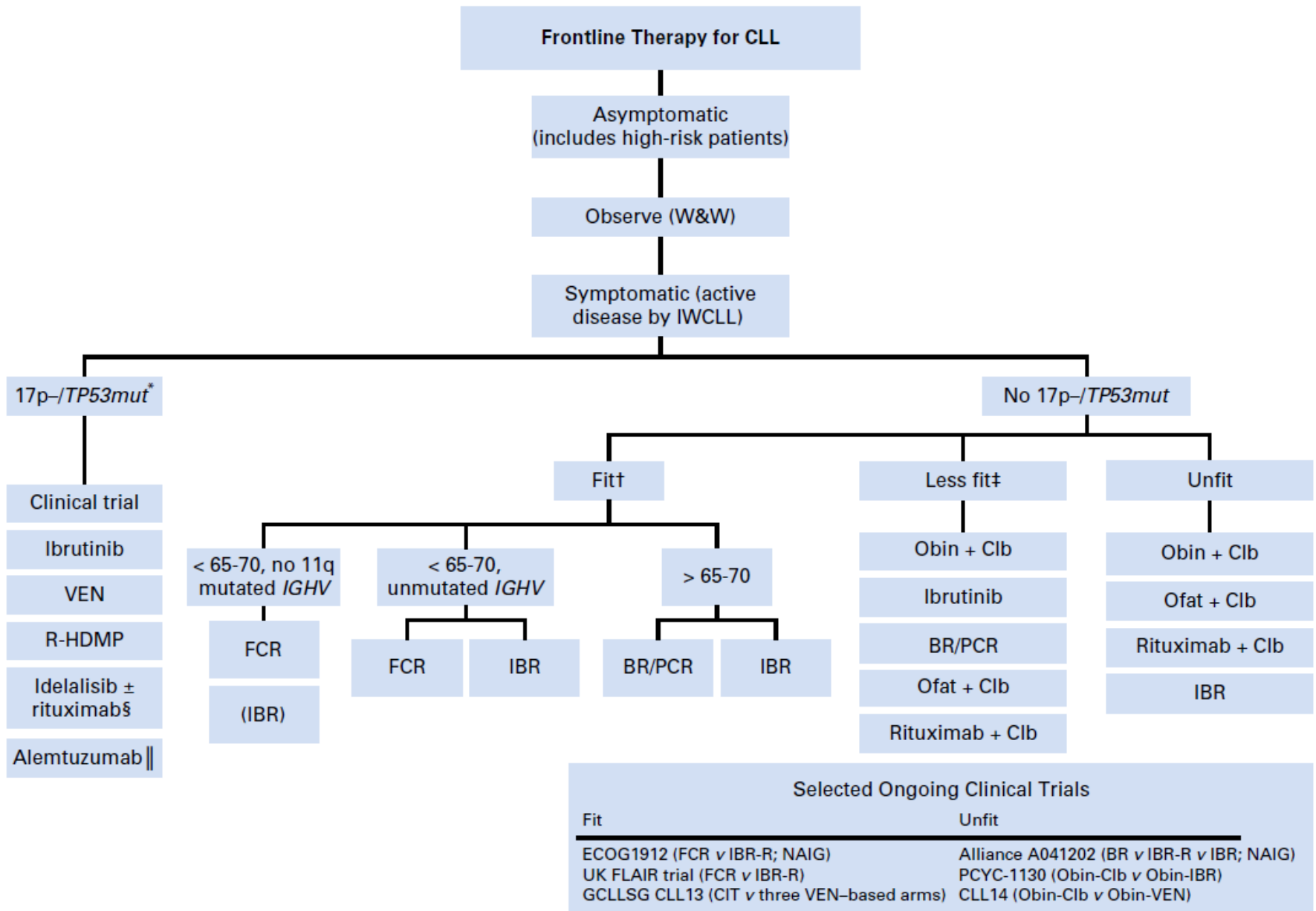
## Best Percent Change from Baseline in Nodal Size by CT Scan

### Median Time to 50% Reduction



N = 51 evaluable (at minimum, 6 week assessment).

# Chemoimmunotherapy Is Not Dead Yet in CLL





## Gruppo

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# Domande

Cariotipo: si o no nella pratica clinica prima della terapia?

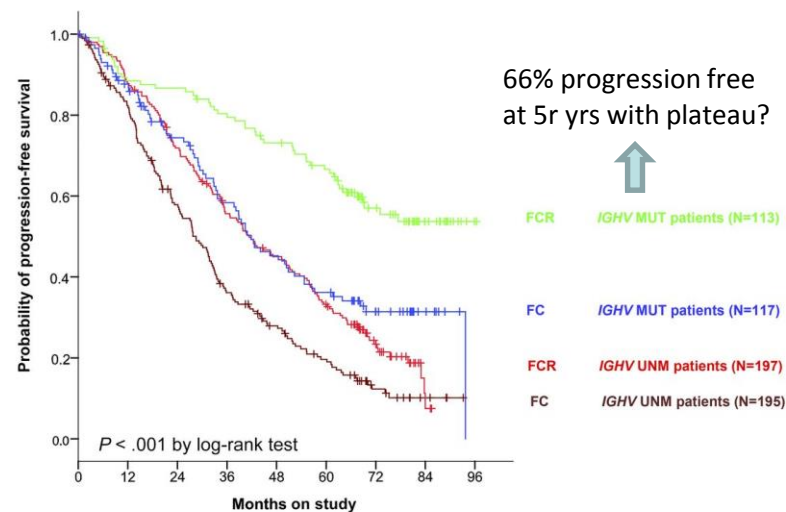
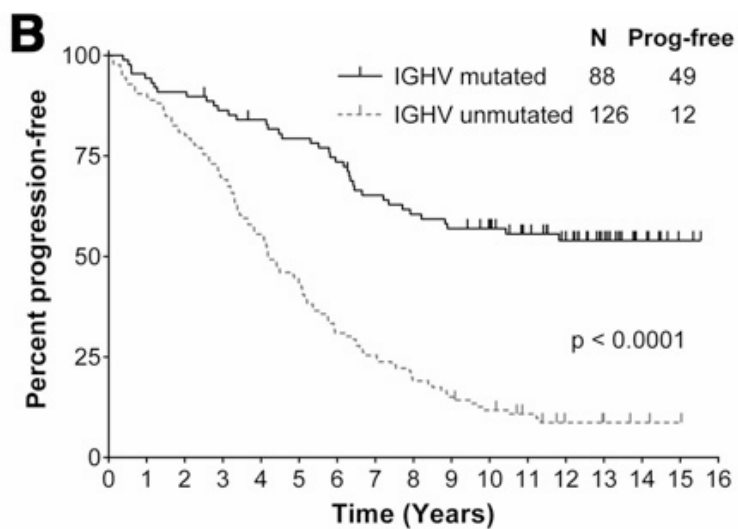
- 8/10 si, in alcuni pazienti
- Significato prognostico da validare
- Alcuni lo eseguono per studi traslazionali
- Non ha ricadute terapeutiche al momento
- Necessità di definire esattamente cosa è il cariotipo complesso

# Domande

**Quale è il vero «bisogno» di miglioramento nella terapia della LLC?**

- 10-15% dei pazienti trattati ha l'eliggibilità a FCR ed è IGHV mutato
- Gli studi dovrebbero trovare terreno ideale nei pazienti a cattiva prognosi

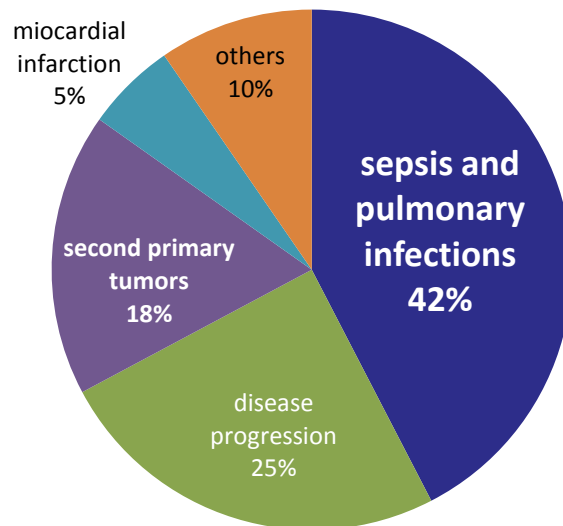
# Long term PFS with FCR (MDACC and GCLLSG – CLL8)



Number at risk	0	12	24	36	48	60	72	84	96
FCR IGHV MUT	113	99	97	89	80	71	37	15	1
FC IGHV MUT	117	96	75	58	45	36	21	7	0
FCR IGHV UNM	197	173	140	106	85	61	25	2	0
FC IGHV UNM	195	153	105	65	45	30	12	4	0

# Causes of death after FCR in the CLL8 trial

FCR arm (n.125 events / 408 patients; 5,9 yrs median f.u.)



## Median time to onset (months) after last dose of study treatment

sepsis and pulmonary infections	46
second primary tumors	27

## Median PFS in high risk CLL treated by Chlor + anti CD20 (elderly/unfit)

	<b>11q-</b>	<b>No 11q-</b>	<b>Unmutated IGHV</b>
<b>Treatment</b>	<b>Chlor + R (UK trial)<sup>1</sup></b>		<b>Chlor + R (GIMEMA trial)<sup>2</sup></b>
<b>Median TTP or PFS (months)</b>	<b>12</b>	<b>24</b>	<b>22,8</b>

1. Hillmen P et al, J Clin Oncol. 2014 Apr 20;32(12):1236-41

2. Foà R et al. Am J Hematol. 2014 May;89(5):480-6

# Domande

**MRD- è un endpoint importante?**

- Sicuramente si nei trials clinici
- In chi ha prognosi buona (IGHV mutato) FCR va bene.
- Utilizzare nuove combinazioni
  - nel salvataggio
  - in subset a cattiva prognosi in prima linea

# Domande

## Ibrutinib in prima linea >65 a chi?

- Solo IGHV non mutato (risponde di più in vitro)
- Tra gli IGHV non mutati: sulla base dell'età
- Quasi mai perché funziona in seconda e terza
- <10-25% ; uno solo >50%



# Possibility to cross in case of discontinuation in rel/ref CLL (toxicity or progression)

## **Ibrutinib**

*18-28% discontinuation at 1 yr  
in the clinical practice*

## **Idelalisib**

*77% discontinuation  
in 3 trials*

## **Venetoclax**

*40% discontinuation  
In trials*

Mato A et al, ASH 2015 oral abs #719  
Coutre S et al, EHA 2016 abs #223

Tam et al  
ASH 2015 poster abs#2939

Idelalisib

Ibrutinib

venetoclax

Better than chemoimmunotherapy

# Domande

## Failure of a KI

Se progressione - venetoclax (MRD-); ibrutinib, idelalisib

Se tossicità: switch ad altro KI